Detecting water stress effects on fruit quality in orchards with time-series PRI airborne imagery

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A B S T R A C T
A methodology for the assessment of fruit quality in crops subjected to different irrigation regimes is presented. High spatial resolution multispectral and thermal airborne imagery were used to monitor crown temperature and the Photochemical Reflectance Index (PRI) over three commercial orchards comprising peach, nectarine and orange fruit trees during 2008. Irrigation regimes included sustained and regulated deficit irrigation strategies, leading to high variability of fruit quality at harvest. Stem water potential was used to monitor individual tree water status on each study site. Leaf samples were collected for destructive sampling of xanthophyll pigments to assess the relationship between the xanthophyll epoxidation state (EPS) and PRI at leaf and airborne-canopy level. At harvest, fruit size, Total Soluble Solids (TSS) and Tritatable Acidity (TA) were measured to characterize fruit quality. A statistically significant relationship between EPS and PRI was found at the leaf (r² = 0.81) and canopy level (r² = 0.41). Airborne-derived crown PRI calculated from the imagery acquired during the fruit growth was related to the ratio of the total soluble solids normalized by the titratable acidity (TSS/TA), an indicator of fruit quality measured on the same trees, yielding a coefficient of determination of r² = 0.50. The relation between the integral of PRI time-series and TSS/TA yielded a coefficient of determination of r² = 0.72 (peach) and r² = 0.61 (nectarines). On the contrary, the relation between TSS/TA and the time-series of crown thermal imagery was very weak (r² = 0.21 and 0.25 respectively). These results suggest that a physiological remote sensing indicator related to photosynthesis, such as PRI, is more appropriate for fruit quality assessment than crown temperature, the established method of water stress detection, which is more related to crown transpiration. A radiative transfer modelling study was conducted to assess the potential validity of this methodology for fruit quality assessment when using medium spatial resolution imagery. The analysis shows important effects of soil and shadows on the PRI vs EPS relationship used for fruit quality assessment if non-pure crown reflectance was extracted from the imagery.

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1. Introduction

Twenty-five years ago, thermal information was chosen for the remote sensing of water stress in crops (Jackson et al., 1981; Idso, 1982a,b) because the spectral vegetation indices that existed at that time were not nearly as sensitive to water deficits as those derived from canopy temperature (Jackson et al., 1983). Thermal remote sensing of water stress was first performed using spectrometers at ground level (Idso et al., 1981; Jackson et al., 1977, 1981), but other approaches have been developed more recently. These included the use of airborne thermal imagery (Cohen et al., 2005; Leinonen & Jones, 2004; Sepulcre-Cantó et al., 2007) and satellite thermal information in combination with 3D radiative transfer models to understand the effects of scene thermal components on large ASTER pixels (Sepulcre-Cantó et al., 2009). Notwithstanding the advances in thermal detection, the visible part of the spectrum has also been useful for pre-visual water stress detection based on indices that use bands located at specific wavelengths where photosynthetic pigments are affected by stress condition. This is the case of the Photochemical Reflectance Index (PRI) (Gamon et al., 1992) that has been proposed to assess vegetation water stress based on xanthophyll composition changes (Peguero-Pina et al., 2008; Suárez et al., 2008, 2009; Thenot et al., 2002). The PRI was presented as an indicator of the epoxidation state of the xanthophylls pool or, what is the same, the proportion of violaxanthin that has been converted into zeaxanthin under stress conditions (Gamon et al., 1992). For water stress detection, PRI could be an alternative to thermal remote sensing, enabling the use of low-cost imaging sensors with high spatial resolution capabilities that are not possible in the thermal domain (Suárez et al., 2008, 2009).

In addition, the PRI is an index that was first formulated as an indicator of photosynthetic efficiency, but is also an indicator of
photosynthesis rate through light use efficiency (Asner et al., 2005; Drolet et al., 2005; Fuentes et al., 2006; Guo & Trotter, 2004; Nakaji et al., 2006; Nichol et al., 2000, 2002; Serrano & Peñuelas, 2005; Sims et al., 2006; Strachan et al., 2002; Trotter et al., 2002) and through chlorophyll fluorescence (Dobrowsky et al., 2005; Evin et al., 2004; Nichol et al., 2006). Therefore, PRI in addition to being a water stress indicator, is also directly related to several physiological processes involved in the photosynthetic system.

The remote detection and monitoring of water stress is critical in many world areas where water scarcity is a major constraint to irrigated agriculture, and is forcing farmers to reduce irrigation water use via deficit irrigation (DI) (Fereres & Soriano, 2007). One of the DI approaches is the regulated deficit irrigation (RDI), where water deficits are imposed only during the crop developmental stages that are the least sensitive to water stress (Chalmers et al., 1981). This practice was originally proposed to control the vegetative vigour in high-density orchards to reduce production costs and to improve fruit quality. However, it also saves irrigation water, with the concomitant benefits of reduced drainage losses (Fereres & Soriano, 2007). It has long been known that tree water deficits affect fruit quality parameters (Veihmeyer, 1927). However, when water deficits are imposed as in RDI, yield and fruit size are not affected (Girona, 2002), while some quality parameters such as total soluble sugars and total acidity increase (Crisosto et al., 1994; Girona et al., 2003; Mills et al., 1994). The responses to RDI are variable depending on the timing and severity of the RDI treatments that are the least sensitive to water stress (Marsal & Girona, 1997; Girona et al., 2003) which vary within a given orchard; thus the need for remote sensing tools that could assist in monitoring stress over entire orchards. Additionally, the changes in irrigation depths with time and the lack of uniformity in water application during the irrigation period emphasize the need for a methodology that would cover the entire season, integrating the shortterm variations in tree water status. One option would be to use an integrated measure over time of tree water status (Myers, 1988; Ginestar & Castel, 1996). González-Altozano and Castel (1999) related the time integral of stem water potential with yield and fruit quality parameters in citrus. Baeza et al. (2007) attempted the same approach on vineyards, finding a correlation between a water stress-integral and final berry size, although not with sugar composition. Although the relationships between water stress and fruit quality has been widely studied, the conclusion is that there is a lack of reliable indicators that predict with precision final fruit quality, and therefore there is a need for further research concerning potential fruit quality indicators.

Remote sensing of fruit quality has been attempted by several means such as by determining the vigour or total leaf area in vineyards (Johnson et al., 2001, 2003; Lamb et al., 2004); by relating quality parameters in water-stressed mandarin trees to spectral changes in the red and green channels (Kroon-Vizzi et al., 2008), and by using high spatial resolution airborne thermal imagery to outline relationships of olive fruit size, weight, and oil content against thermal water stress indicators (Sepulcre-Cantó et al., 2007).

In this work, the PRI has been used to assess fruit quality parameters in peach and orange orchards under various water regimes. A time-series of airborne PRI imagery over a peach and an orange orchard under different irrigation treatments were acquired and related to fruit quality at harvest. Furthermore, a 3D radiative transfer model was used to assess the applicability of this method to medium resolution PRI imagery for extended monitoring of crops at larger scales. For this purpose, simulations using different soil backgrounds were conducted and the output spectral information was evaluated at different spatial resolutions.

2. Material and methods

2.1. Study sites

The experimental areas are located in Western Andalucía, Spain, a region of Mediterranean climate characterized by warm and dry summers and cool and wet winters, with an average annual rainfall of over 550 mm.

The first study site was located on a commercial peach orchard planted in 1990 in a 5 × 3.3 m grid on a deep soil with moderately high water holding capacity and classified as Typic Xerofluvent in Cordoba, Spain (37.5°N, 4.9°W) (Fig. 1a). Two experiments were carried out in this location. One experiment was conducted using a set of eight rows of nine peach trees (Prunus persica cv. “BabyGold 8”). Within that set, 18 trees were drip irrigated starting in mid May (end of Stage I of fruit growth and beginning of Stage II) at a rate that met the evapotranspiration (ET) requirements (full-irrigated treatment or control). Additionally, three different RDI treatments were applied to plots of 12 trees each, by varying the onset of re-irrigation as Stage III of fruit growth commenced, following uniform water deficits imposed in Stage II. The dates of onset of re-irrigation (at 160% of ET) were 4 July, 11 July and 17 July, respectively. In the same commercial orchard, another experiment was conducted on nectarine trees (Prunus persica cv. “Sweet Lady”). Five rows of 30 trees each were irrigated meeting the ET requirement following the commercial orchard schedule, while another six rows of 30 trees each were subjected to an RDI regime that imposed water deficits until 30 June, the beginning of Stage III (Fig. 1b).

The second study site was located near Seville, Southern Spain (37°N, 5.7°W), on an 82 ha commercial citrus orchard (Fig. 1c shows the subset where the experiment was established). The light-textured soil is classified as Rodoxertox, with an approximate depth of 3 m. The trees were planted in 1997 in a 7 × 3 m pattern on a N–S orientation. The experiment was a randomized block design with six replications, each individual plot composed of five rows of three orange trees (Citrus sinensis L. cv. “Navelina”). Three different drip irrigation treatments were applied: i) the control, that followed the orchard schedule which is designed to meet ET requirements for maximum production; ii) an over-irrigated treatment that applied 137% of ET; and iii) a DI treatment that applied 62% of control. Table 1 lists the treatments, irrigation periods and depths, harvest and imagery acquisition dates for each of the study sites.

2.2. Field data

From the beginning of June till harvest, stomatal conductance (G) and stem water potential (Ψ) were measured weekly at midday with a leaf porometer (model SC-1, Decagon Devices, Washington, DC, USA) and a pressure chamber (PWS-C Model 3000, Soil Moisture Equipment Corp., California), respectively. For the first experimental site, measured Ψ values for the three RDI treatments were normalized by dividing them into the control Ψ values of the full-irrigated treatment.

Leaf spectral measurements of adaxial surfaces were taken with a leaf probe that was attached to a field spectrometer ASD measuring in the spectral range of 400–1100 nm with a 25° field of view (FieldSpec Handheld Pro, ASD Inc., CO, USA). Measurements were taken on four leaves per crown, which were collected and immediately frozen in liquid nitrogen. For these, leaf spectral properties and indices were calculated to assess relationships with pigment concentrations determined after extraction through chromatography (see below).

Two square centimeters of leaf tissue were obtained with a cork borer from each of the four leaves per tree crown. The discs were frozen in liquid nitrogen in the field and later kept under −20 °C in microfuge tubes. Each leaf disc set corresponding to each crown was ground in a mortar on ice with liquid nitrogen and aceton (in the presence of Na ascorbate) up to 5 ml. Then, the extract was filtered through a 0.45 µm filter to separate vegetation residues and Na ascorbate and left in dark tubes at −20 °C for pigment analysis, as reported by Abadía and Abadía (1993). First, absorption at 470, 644.8 and 661.8 nm was measured with a spectrophotometer in order to derive chlorophyll a and b, and total carotenoid concentrations as
described by Abadía and Abadía (1993). Samples were injected in a 100×8 mm Waters Novapak C18 radial compression column (4 µm particle size) with a 20 µl loop and mobile phases were pumped by a Waters M45 high pressure pump at a flow of 1.7 ml/min (Larbi et al., 2004).

In all three orchards, between 12 and 36 trees located in the centre of the irrigation treatments were monitored and harvested individually. All fruits from each tree were immediately weighted and their diameters measured. Later, eight fruits per tree were selected randomly for a physiochemical and organoleptic characterization of Total Soluble Solids (TSS) and Total Acidity (TA) used to calculate the ratio (TSS/TA). The combination of TSS and TA in the ratio TSS/TA is an indicator of both sweetness and fruit acidity, giving more information than TSS or TA separately (Crisosto et al., 2006). Fruit juice was obtained, filtered and measured with a pH meter (pH-Burette 24, Crison, Spain). To measure Titratible Acidity, 6ml of juice from each sample were mixed with 50 ml of water and used for titrating with 0.1 N NaOH to an end point of pH 8.2. The total volume of NaOH is

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**Fig. 1.** Overview of the experiments conducted on peach trees (a); nectarines (b); and orange trees (c). The irrigation treatments for the peach study area consisted of fully-irrigated and three deficit irrigation treatments (RDI1, RDI2 and RDI3). The nectarine field was divided into one fully-irrigated treatment and five rows of trees under regulated deficit irrigation (RDI). In the orange tree experiment there were three different irrigation treatments: over-irrigated, control and deficit-irrigated.
measured and used to calculate the Titratable Acidity using the Eq. (1).

\[
\% \text{ Acid} = \frac{\text{ml(NaOH)} \times (0.1 \text{N NaOH}) \times (\text{milliequivalent factor}) \times 100}{\text{grams of sample}}
\]  

(1)

The milliequivalent factor is a coefficient dependant on the most predominant acid in the fruit. In the case of peach trees, malic acid is predominant and the milliequivalent factor is 0.067. For orange trees, citric acid is predominant, with a milliequivalent factor of 0.064. Soluble solids concentration (SSC, °Brix) was determined in a small sample of fruit juice using a hand-held refractometer (Atago, ATC-1E, Japan).

Pigment concentrations were derived from the total area of the peaks in the chromatogram using previously determined calibration coefficients calculated by injecting pure pigments into the HPLC circuit. From the xanthophyll pigment concentrations of violaxanthin (V), antheraxanthin (A), and zeaxanthin (Z), the epoxidation state (EPS) was calculated as \((V + 0.5A)/(V + A + Z)\) (Thayer & Björkman, 1990). The EPS was calculated from the leaf set corresponding to each crown and compared with the PRI calculated as \((R570 - R531)/(R670 + R531)\) (Gamon et al., 1992), from the spectra measured in the field on the same four leaves sampled.

2.3. Airborne imagery

A 6-band multispectral camera (MCA-6, Tetracam, Inc., California, USA) was flown in the summer of 2008 at 150 m above the ground level using an unmanned aerial vehicle (UAV) (Berni et al., 2009), acquiring a time-series of 12 images on different dates from the two study sites. Eleven images were acquired on the commercial field covering the nectarine and peach experiments from the beginning of the Stage II of fruit growth (12th of June) till the end of harvest (21 August). In the orange orchard, the water status of the different irrigation treatments was kept constant. Hence, there was no need for characterizing the water stress over the whole fruit growing period, and a single image acquired on 16 September was used. The camera has six image sensors with 25 mm diameter bandpass filters of 10 nm FWHM (Andover Corporation, NH, USA). The image resolution is 1280 × 1024 pixels with 10-bit radiometric resolution and optical focal length of 8.5 mm, yielding a ground-based spatial resolution of 15 cm at 150 m altitude. The bandsets used in each of the study sites included those centered at 530 and 570 nm used to calculate the PRI index, as well as 550, 670, 700 and 800 nm to calculate the TCARI/OSAVI index for chlorophyll content estimation (Haboudane et al., 2002), and the NDVI (Rouse et al., 1974), and the SR for LAI estimation. An overview of the spectral indices used in this study can be found in Table 2. Geometric calibration of airborne data was conducted as explained in Berni et al. (2009). The camera was radiometrically calibrated using coefficients derived from measurements made with a uniform calibration body (integrating sphere, CSTM-USS-2000C Uniform Source System, LabSphere, NH, USA) at four different levels of illumination and six different integration times. Radiance values were later converted to reflectance using total incoming irradiance simulated using sunphotometer (Microtops, Solar Light Inc.) data taken in the field at the time of imagery acquisition.

A thermal camera (Thermovision A40M; FLIR, USA) was installed onboard the airborne platform. Its image resolution was 320 × 240 pixels and 16bits of at-sensor calibrated radiance with a 40° FOV lens, yielding 40 cm spatial resolution at 150 m altitude. The image sensor is a Focal Plane Array (FPA) based on uncooled microbolometers with a spectral range of 7.5–13 μm, yielding calibrated radiance in the range of 233–393 K. The methodology for obtaining surface temperature from radiance temperature by removing atmospheric effects using a single-channel atmospheric correction is explained in Berni et al. (2009).

As PRI is an index related to light use efficiency, the index was normalized with the incoming PAR over the time-series at the time of image acquisition, calculated as the integral of the irradiance in the visible region of the electromagnetic spectrum (range 400–700 nm). Thus, PRI/PAR values could be used to study the time-series of the stages II and III of fruit growth. Reflectance values were obtained for the six spectral bands for every single crown for the whole time-series. The high spatial resolution allowed the identification of each

| Table 1 |
| Summary of each study site, harvest, and imagery acquisition dates. |
| Species | Treatments | Irrigation strategy | Withheld period | Irrigation dose | Harvest Flights |
| Nectarine | Control | Sustained | 05/21 to 06/30 | 100% ET | From 07/15 to 08/07 |
| Peach | RDI-1 | Regulated | 05/19 to 07/04 | Re-watering: 200% ET, later: 100% ET | 06/12, 06/19, 06/25, 07/01, 07/04, 07/17, 07/23, 07/29, 08/07, 08/12, 08/16 |
| Peach | RDI-2 | Regulated | 05/19 to 07/11 | Re-watering: 160% ET, later: 80% ET | 06/19, 06/25, 07/01, 07/04, 07/17, 07/23, 07/29, 08/07, 08/12, 08/21 |
| Peach | RDI-3 | Regulated | 05/19 to 07/18 | Re-watering: 160% ET, later: 80% ET | 08/06, 08/28 |
| Peach | Over-irrigated | Sustained | 100–130% ET | 11/03 |
| Orange | Control | Sustained | 06/12, 06/19, 06/25, 07/01, 07/04, 07/17, 07/23, 07/29, 08/07, 08/12, 08/16 |
| Orange | Over-irrigated | Sustained | 09/16 |

The treatments under RDI had a "withheld period" (no irrigation) followed by a re-watering period until their stem water potential were equal to the control.

| Table 2 |
| Overview of the vegetation indices used in this study and their formulation, with \( R_\lambda \) being the reflectance at \( \lambda \) nm. |
| Normalized Difference Vegetation Index: \( \text{NDVI} = \frac{R_{\text{red}} - R_{\text{nearIR}}}{R_{\text{red}} + R_{\text{nearIR}}} \) | Rouse et al. (1974) |
| Transformed Chlorophyll Absorption in Reflectance Index/Optimized Soil-Adjusted Vegetation Index: \( \text{TCARI} / \text{OSAVI} = \frac{R_{\text{green}} - 1}{0.5 R_{\text{green}} - R_{\text{red}}} - 0.2 \frac{R_{\text{green}} - R_{\text{red}}}{R_{\text{green}} + R_{\text{red}}} - 0.16 \frac{R_{\text{green}} - R_{\text{red}}}{R_{\text{green}} + R_{\text{red}}} + 0.04 \frac{R_{\text{green}} - R_{\text{red}}}{R_{\text{green}} + R_{\text{red}}} \) | Haboudane et al., 2002 |
| Simple Ratio: \( \text{SR} = \frac{R_{\text{red}}}{R_{\text{nearIR}}} \) | Asrar et al. (1985) |
| Photochemical Reflectance Index: \( \text{PRI} = \frac{R_{\text{green}} - R_{\text{red}} - R_{\text{nearIR}}}{R_{\text{green}} + R_{\text{red}} + R_{\text{nearIR}}} \) | Gamon et al. (1992) |
individual tree and the possibility of selecting pure sunlit vegetation pixels. The pixels at the edge of the crowns were not used to extract the spectral information. Additionally, individual crown temperature \((T_c)\) was extracted from thermal imagery avoiding crown edge pixels and was normalized with air temperature \((T_a)\) to use the value of \(T_c - T_a\) for the analysis. The integral over time was calculated for five indices: PRI/PAR, \(T_c - T_a\), NDVI, SR, and TCARI/OSAVI for the period from 19th of July to 7th of August when the harvest started. For indices having both negative and positive values, the minimum value found for each index along the whole period was used as a reference line to calculate the integral for all the trees, as explained in Myers (1988).

In order to study the effect of the spatial resolution on PRI, the aggregated reflectance including shadows, soil and crown components was extracted for each crown in all images acquired, and the integral calculated. For this purpose, regions of interest comprising each individual crown, and the adjacent soil were created and the spectral information extracted.

2.4. Radiative transfer modelling

The 3D Forest Light interaction radiative transfer model (FLIGHT, North, 1996) was used to investigate the application of this methodology to larger spatial resolution imagery, by assessing canopy aggregation effects on PRI. The FLIGHT model has been successfully used to simulate PRI in previous studies (Barton and North, 2001; Suárez et al., 2008, 2009). In this case, typical peach leaf spectral characteristics and tree structural parameters were inputs for simulating vegetation cover ranging from 10% to 100% on three different soil types. Typical peach leaf reflectance and transmittance spectra were simulated using PROSPECT model (Jacquemoud and Baret, 1990) for \(N = 1.6, C_{ab} = 40 \mu g/cm^2, C_m = C_w = 0.015\) and \(C_s = 0\). The input values of \(N, C_m, C_w,\) and \(C_s\) were found in previous literature on peach trees (Suárez et al., 2009), the input value for \(C_{ab}\) was defined as the average of the chlorophyll concentrations estimated by destructive methods. The canopy input parameters used were crown LAI = 2.5, LAD = spherical and leaf size = 0.02 m. The solar geometry corresponded to mid July at 10:00 GMT in Cordoba, Spain (solar zenith = 23° and solar azimuth = 124°). Fig. 2 represents the three generic soil spectra used (Fig. 2a) and three simulations conducted for dark soil and 30% (Fig. 2b), 50% (Fig. 2c), and 70% vegetation cover (Fig. 2d). From each simulation, crown reflectance and scene reflectance were extracted by digitizing regions of interest on the simulated scene image to calculate crown PRI and scene PRI, respectively. Crown PRI values were compared to scene PRI values in order to assess the effects caused by the spatial resolution on the index. One of the outputs of FLIGHT radiative transfer model is the percentage of each element: shadowed soil, sunlit soil, shadowed vegetation and sunlit vegetation in the whole simulated scene. Those percentages were used to assess the magnitude of the error when deriving EPS from an aggregated pixel of soil, shadows and vegetation as a function of the soil type. For this purpose, scenes with a vegetation cover of 50% were simulated using the soil spectra in Fig. 2. The aggregated pixel reflectance was calculated as the sum of the fractional covers of each element in the scene multiplied by its reflectance. The vegetation fractional cover was considered to be the sum of the fractional covers for sunlit and shadowed vegetation from the FLIGHT output. The aggregated pixel reflectance was calculated as the sum of the fractional covers of each element in the scene multiplied by its reflectance. The vegetation fractional cover was
considered to be the sum of the fractional covers for sunlit and shadowed vegetation from the FLIGHT output. The aggregated scene reflectance was the result of using tree reflectance which corresponded to the vegetation fractional cover in the scene. Three different scenes were recreated per tree using three soil types. For each tree, the EPS value was compared to the aggregated PRI calculated from tree reflectance and the reflectance of the fractions of shadowed and sunlit soil.

3. Results and discussion

Fig. 3a shows that, at the leaf level, the EPS calculated from pigment determination methods was well correlated with leaf PRI calculated from the same leaves collected in the field. Leaves with higher EPS values, corresponding to a high concentration of the photosynthetic active pigment violaxanthin over the whole xanthophyll pool, and consequently less stressed, presented lower PRI values. Lower values of PRI are the consequence of lower absorption at 530 nm using the presented formulation of PRI corresponding to a lower concentration zeaxanthin, which is the xanthophyll pigment inhibiting photosynthesis (Gamon et al., 1992). At the crown level, the photon interaction with vegetation structure and the effect of leaf angle distribution result in lower PRI values as we can see in Fig. 3b and c. These figures depict the relationships obtained between EPS, crown PRI ($r^2 = 0.41$), and crown NDVI ($r^2 = 0.15$) extracted from the canopy images. The data shown on Fig. 3b and c suggest that PRI changes are driven by physiological processes related to xanthophyll pigments and not by vegetation structure. At both leaf and canopy scales, PRI and chlorophyll a+b content were not correlated, yielding coefficients of determination of $r^2 = 0.04$ and $r^2 = 0.01$ respectively (data not shown), indicating that PRI changes were not driven by chlorophyll content differences. Several authors have previously found an inverse relationship between EPS and PRI (Filella et al., 2002).

Fig. 3. (a) Relationship between the epoxidation state (EPS) calculated from xanthophyll pigment extraction and PRI from spectral measurements on the same leaves from a total of 12 tree crowns. Relationship between the EPS calculated from the averaged xanthophyll content of four leaves per crown and spectral indices extracted from imagery for the crowns: PRI index (b) and NDVI (c).

Fig. 4. Correlation between pre-harvest EPS in the nectarine orchard experiment and the fruit quality ratio, Total Soluble Solids/Tritatable Acidity (TSS/TA).

### Table 3

<table>
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<th>Orange</th>
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<th>TA</th>
<th>TSS/TA</th>
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<td></td>
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Pearson's analysis.

* Correlation is significant at the 0.01 level (2-tailed) (In bold).

**Correlation is significant at the 0.05 level (2-tailed).
as shown in Fig. 3b. The results in Fig. 3 demonstrate the link between PRI and the xanthophyll cycle in peach at both leaf and canopy scales.

The different irrigation treatments generated variability in fruit quality, as assessed by the TSS/TA ratio measured at harvest. The TSS/TA variability in the peach orchard with four irrigation treatments was greater than the TSS/TA variability found in the nectarine and orange orchards, where only one deficit irrigation treatment was applied. Fig. 4 shows the relationship between EPS at crown scale and TSS/TA.

Table 4

<table>
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<th>Peach</th>
<th>Nectarines</th>
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</table>

Pearson's analysis.

**Correlation is significant at the 0.01 level (2-tailed) (In bold).

*Correlation is significant at the 0.05 level (2-tailed).
measured at harvest for the nectarine experiment. The higher values of EPS, obtained from trees under full irrigation, corresponded to lower values of TSS/TA found in fruits of that treatment. Previous studies on peach have demonstrated the inverse relationship between tree water status at Stage II of fruit growth and the ratio TSS/TA (calculations from data published by Crisosto et al., 1994; Gelly et al., 2003).

In the orange orchard experiment, one treatment was subjected to sustained but mild deficit irrigation throughout the irrigation season. Measurements of PRI, NDVI, SR, TCARI/OSAVI and temperature extracted from imagery were related to fruit parameters. The relationships of fruit quality parameters (TSS, TA, TSS/TA and fruit size) with the physiological and structural indices are listed in Table 3. PRI showed the highest correlation with TA and TSS/TA, which are considered important indicators of fruit quality. On the contrary, crown temperature was not associated with any of the quality parameters, with one exception (fruit size; $r^2 = 0.47$). This is consistent with the stem water potential measurements, which indicated no significant differences between the deficit irrigation treatment and the control. Consequently, crown temperature differences between the two treatments were hardly detectable. The PRI image on the orange orchard was acquired during the stage of rapid fruit growth. Fig. 5 shows the relationship of TSS/TA with: a) PRI; b) temperature; c) NDVI; and, d) TCARI/OSAVI. Crown PRI correlated reasonably well with TSS/TA ($r^2 = 0.50$), while other indicators of water stress, such as temperature, structural indices such as NDVI and SR, and an index related to chlorophyll content TCARI/OSAVI were not related to TSS/TA, (coefficients of determination of 0, 0.16, 0.17 and 0.18, respectively; Fig. 5 and Table 3). Although canopy temperature is a reliable remote sensing water stress indicator (Jackson et al., 1977), crown temperature values should be related to the instantaneous transpiration rate at the time of image acquisition. By contrast, crown PRI values reflect physiological processes related to photosynthesis as affected by water stress. Because fruit quality is more tied to photosynthesis and carbon metabolism, PRI may be a better estimator of fruit quality than other established water stress indicators that are related directly to transpiration such as crown temperature.

In the peach experiment, the time courses of water deficits and recovery were monitored by measuring crown PRI from imagery and SWP in the field. The water-stressed trees showed lower SWP and higher crown PRI values (representing stress conditions) than the control trees. Fig. 6 shows the time-series of PRI/PAR and the normalized SWP values (values divided by those of the control for Stages II and III of fruit growth versus the fruit quality ratio, TSS/TA for peach (a and c) and nectarine (b and d).
During the water deficit period that corresponded to Stage II of fruit growth, the values of PRI/PAR for the three deficit irrigation treatments were similar and higher than the control values, indicating water stress (Fig. 6). Similarly, the SWP of those treatments were lower than that of the fully-irrigated treatment. As the rapid growth or stage III of fruit growth began in 4 July (“Irrigation 1”), the RDI-1 treatment was re-irrigated until its SWP recovered to control values, a process that took 5 days. One week later (“Irrigation 2”), the RDI-2 treatment was re-irrigated, and the PRI/PAR for both RDI-1 and RDI-2 treatments fell below the control values (Fig. 6) showing recovery of water status. Equally, SWP values recovered in RDI-1 and RDI-2 until they reached the values of the fully-irrigated treatment, although the RDI-2 treatment took ten days to recover (Fig. 6). The RDI-3 treatment entered the recovery phase at “Irrigation 3”; both the SWP and PRI/PAR records showed that recovery in this treatment occurred after 13 days, taking longer to recover from the more severe water stress than the other two RDI treatments. The results of Fig. 6 clearly demonstrate that the PRI/PAR measurements tracked the evolution of tree water status in the various treatments.

In the peach experiment, the variation in irrigation regimes with time indicated that a single measurement of PRI could not glean stress history. Therefore, it was necessary to use a time-series of data acquired during fruit growth to accurately describe the different treatments. Table 4 presents the results of analyzing the indices time-series (computed over Stages II and III of peach and nectarine fruit growth) against the parameters TSS, TA, TSS/TA and fruit size. Again, the ratio TSS/TA was best correlated with the integral of PRI/PAR for peach and nectarine trees. Fig. 7 shows an independent analysis of the two water stress indicators: PRI/PAR and $T_c - T_a$. Fig. 7a and c show the relationship of the integral of PRI/PAR and $T_c - T_a$ with TSS/TA for
peach, and Fig. 7b and d show the same for nectarines. The correlation of TSS/TA with the integral of PRI/PAR calculated for every single crown using a set of ten images taken in different days yielded linear relationships of $r^2 = 0.72$ for the peach tree experiment, and $r^2 = 0.61$ for the nectarine orchard (Fig. 7a and b, respectively). By contrast, relationships between the integral of $T_a - T_T$ with TSS/TA, yielded much lower linear relationships for peach ($r^2 = 0.21$) and nectarines ($r^2 = 0.25$) (Fig. 7c and d). A possible explanation for the difference in behaviour between PRI and $T_a - T_T$ lies in the changes in carbon partitioning in response to mild water deficits. It has been shown that deficit irrigation alters the distribution of carbon, increasing the allocation to fruits (Fereres and Soriano, 2007). Mild water deficits that hardly would affect transpiration (and hence $T_a - T_T$) may have some effects on carbon metabolism that are reflected in the light reactions of photosynthesis and are detected by PRI measurements.

The effect of water stress on fruit size, an important commercial parameter (Guardiola & García-Luis, 2000), was studied in peach and nectarine using multispectral imagery acquired during Stage III of exponential fruit growth, when the absence of water stress is critical (Génard & Huguet, 1996). PRI/PAR time-series computed over Stage III of fruit growth correlated well with fruit size at harvest ($r^2 = 0.51$, data not shown), while the correlation of $T_a - T_T$ time-series yielded a much smaller coefficient of determination ($r^2 = 0.15$). Fig. 8 presents an overview of the peach experiment with the different treatments and a zoom on four of the monitored trees, each of them corresponding to a different irrigation schedule: Full-irrigated, RDI–1, RDI–2 and RDI–3 (Fig. 8a). Fig. 8b to e show the ratio TSS/TA for the four trees. In Fig. 8f to i, the integral of PRI/PAR is shown for each of the selected and surrounding trees, demonstrating that the variability among trees of the water stress-integral within-treatments may be detected using an image-based methodology. The values of the integral of PRI/PAR and TSS/TA represented using the same color code, appear the same for the four selected trees, demonstrating that the detection of fruit quality is possible using the time-series of PRI/PAR. Moreover, the use of remote sensing for the assessment of fruit quality parameters permits the spatial characterization of an entire orchard.

The assessment of the influence of the imagery spatial resolution on PRI was studied using image data and radiative transfer modelling. The performance of the integral of PRI/PAR for different spatial resolutions was assessed using data from imagery. First, the reflectance extracted from pure crowns was used (Fig. 9a), and then the reflectance extracted from aggregated pixels, including soil, shadows and crowns (Fig. 9b). The use of high spatial resolution imagery (zoom shown in Fig. 9c) allowed the classification of the crown into sunlit vegetation pixels and shadowed vegetation pixels. Fig. 9d presents an example of a supervised maximum likelihood classification in which within-crown sunlit vegetation and shadows can be identified. When the integral of PRI/PAR is calculated using pure crown reflectance (Fig. 10), the integral at crown scale, as a function of the EPS along the fruit growth, is well correlated with the TSS/TA ratio ($r^2 = 0.72$, Fig. 10a). The integral of PRI/PAR extracted from pure crown pixels (i.e., high spatial resolution) versus the integral of PRI/PAR extracted from aggregated pixels where there is crown, soil and shadows (low-spatial resolution) yielded a very low association ($r^2 = 0.06$, Fig. 10b). The lack of relationship when scene components are aggregated in a mixed pixel suggests that the PRI vs. EPS relationship is lost when pure sunlit crowns are not selected. In fact, the relationship of the integral of “low-spatial resolution” PRI with TSS/TA yielded a coefficient of determination of 0.25 (Fig. 10c), versus the $r^2 = 0.72$ when using high spatial resolution PRI. The results of modelling PRI for pure crown and aggregated pixels suggest a large influence of soil and shadows on the index (Fig. 11), which prevents detecting the physiological responses to mild water stress levels. Aggregated PRI values extracted from the full reflectance scene differ greatly from pure crown PRI values as vegetation coverage decreases, increasing the proportion of soil and shadowed background in the aggregated pixel from 100% to 10% vegetation cover. The effects on the index as a function of three soil types (spectra from Fig. 2) were assessed. Dark soils showed a higher influence on aggregated PRI values (see the separation from the 1:1 line), even for high percentage vegetation coverage (Fig. 11). The dark soil spectrum is low and flat in comparison with the two other soil spectra used, then the influence on PRI is higher than the influence of soils with steeper spectra. The possible errors using low-resolution imagery are presented in Fig. 12. The error bars in Fig. 12b represent the variability of the PRI value for an aggregated pixel depending on the soil type. These modelling results emphasize the need for high spatial resolution imagery to reproduce the results obtained in this study, as medium resolution PRI imagery would be heavily affected by background and shadow components, greatly reducing the sensitivity to physiological indicators of stress, such as PRI. Future developments in this field may focus on minimizing background effects in order to use our methodology with lower spatial resolution imagery. One approach could be the use of a soil-adjusted vegetation index to deal with variable backgrounds. The first soil-adjusted vegetation index presented was SAVI (Huete, 1988). Subsequently, additional indices have been proposed such as TSAVI (Baret & Guyot, 1991), MSAVI (Qi et al., 1994), OSAVI (Rondeaux et al., 1996), and GESAVI (Gillabert et al., 2002), among others. Other authors have successfully transformed indices using soil-adjusted lines to minimize soil influence on spectral reflectance (Haboudane et al., 2002). Use of
Fig. 11. Pure crown PRI values extracted from radiative transfer simulation using the 3D forest light interaction model (FLIGHT) versus aggregated PRI including crown, soil and shadows for vegetation cover ranging from 100% to 10% for three backgrounds. Leaf input parameters were $N = 1.6$, $C_{ab} = 40 \mu g/cm^2$, $C_w = 0.015$, $C_m = 0.015$ and $C_s = 0$, canopy input parameters were LAD = spherical, crown LAI = 2.5, solar zenith = 23° and solar azimuth = 124°.

Fig. 12. (a) Relationship between EPS and pure crown PRI for peach trees; (b) relationship between EPS and PRI from aggregated pixels including different backgrounds. The error bars are function of the maximum and minimum scene PRI corresponding to each EPS value.
the above-mentioned approaches may adapt this methodology to lower spatial resolution imagery.

4. Conclusions

This study demonstrates the link between the epoxidation state of the xanthophyll cycle and the fruit quality measured in orchards under different irrigation regimes, enabling the remote detection of fruit quality as a function of water stress using high-resolution airborne PRI. The PRI index measured at leaf scale was in agreement with the epoxidation state of the xanthophyll cycle calculated from destructive sampling. In the airborne, airborne-derived PRI values calculated from pure crown reflectance were also in agreement with xanthophyll EPS measured on the same trees. Moreover, the time-series of airborne PRI normalized by the incoming PAR at the time of imagery acquisition (PRI/PAR) matched the changes in tree water status as affected by the different irrigation regimes. In an orange orchard under sustained deficit irrigation, crown PRI calculated from the airborne imagery acquired during the fruit growth was well related to fruit quality. For peach and nectarine orchards subjected to period of water stress and recovery a methodology was applied that used the time-series of PRI/PAR to assess the fruit quality responses of the different stressed treatments.

Although the high-resolution PRI time-series exhibited a good relationship with fruit quality, crown airborne temperature acquired over the same trees, the established method for remote sensing of water stress, did not yield comparable results and did not correlate with fruit quality. This highlights the advantage of the PRI as a water stress indicator related to other physiological processes and not only to transpiration, for the assessment of fruit quality. Finally, the use of high spatial resolution imagery appears critical for a tight relationship between PRI and EPS, and therefore for assessing fruit quality from airborne PRI measurements. A radiative transfer simulation study demonstrated the influence of soil and shadows on canopy reflectance used to calculate PRI on aggregated pixels, suggesting the critical effects that soil variability has in the computation of the PRI index in discontinuous orchard canopies. This work demonstrates the feasibility for assessing fruit quality in orchards using PRI when high spatial resolution remote sensing imagery is used (20 cm in the present study) as opposed to using lower spatial resolutions where the pixels represent a mixture of shadows, vegetation and soil. The practical implications of this approach to optimize harvest operations and maximize revenues in horticultural crops based on field segmentation as a function of fruit quality may be substantial.

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References


