LEAF NITROGEN ASSESSMENT WITH ISS DESIS IMAGING SPECTROMETER AS COMPARED TO HIGH-RESOLUTION AIRBORNE HYPERSPECTRAL IMAGERY

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ABSTRACT

Traditional methods to estimate leaf nitrogen (N) from satellite imagery rely on structural and chlorophyll \( a+b \) \( (C_{ab}) \) vegetation indices. Recent progress with airborne hyperspectral imagery identified \( C_{ab} \) and SIF as critical indicators for evaluating leaf N variability, yielding superior performance than standard vegetation indices. In tree orchards, accurate physiological assessments require high-spatial-resolution hyperspectral imagery to minimize canopy architecture and soil background effects. Understanding the potential of coarse-spatial-resolution spaceborne hyperspectral imagery for leaf N estimation is critical. In this study, DESIS hyperspectral imagery collected on board the International Space Station was used to assess the quantification of leaf N, evaluating the relative contributions of physiological plant traits and SIF. High-resolution airborne hyperspectral imagery and ground N data were used for validation. Results show that \( C_{ab} \) and SIF were the most critical parameters explaining leaf N both from DESIS and from airborne hyperspectral imagery, yielding strong correlations against ground truth N data \( (r^2=0.90, p<0.0001) \) and with airborne-predicted N \( (r^2=0.75, p<0.0001) \).

Index Terms – DESIS, Nitrogen, Hyperspectral, Spaceborne, Almond, Chlorophyll, SIF, Plant traits

1. INTRODUCTION

Accurate leaf nitrogen (N) assessment is crucial for ensuring adequate nutrient levels and determining fertilizer requirements over the course of the growing season. Monitoring leaf N status at large scales requires remote sensing technologies to achieve affordable quantifications compared with traditional biochemical analyses of leaf tissues. Standard remote sensing methods for N assessment typically use structural and chlorophyll-sensitive vegetation indices derived from multispectral sensors, but relationships saturate at high N levels [1]. Spaceborne hyperspectral sensors can measure detailed spectral features over large areas. The German Aerospace Center (DLR) Earth Sensing Imaging Spectrometer (DEsis) onboard the International Space Station (ISS) is capable of collecting hyperspectral imagery from space. However, leaf N assessment in row-structured orchards is affected by the canopy architecture and background. Thus, the implications of using coarser spatial resolution hyperspectral imagery on the accuracy of N estimation in heterogeneous orchards are crucial.

Previous work [2, 3] on the assessment of leaf N from airborne hyperspectral imagery demonstrated that Solar-Induced Fluorescence (SIF) and physiological plant traits (i.e., \( C_{ab} \) and other leaf biochemicals) retrieved from radiative transfer models (RTM) yielded superior N estimates than standard vegetation indices. These methods showed that \( C_{ab} \) and SIF predicted 95% of the N variability in almond orchards. However, the importance of specific plant traits may differ considerably as coarser spatial resolution due to the structural and background effects. This study investigates the contribution of SIF and the leaf biochemistry quantified by RTM inversions from ISS DESIS hyperspectral imagery for large-scale N assessment in almond orchards, with comparison against high-resolution airborne hyperspectral imagery and ground truth data used as validation.

2. MATERIALS AND METHODS

2.1. Study area

The study site is a commercial almond orchard covering about 1,200 hectares, located in northwestern Victoria,
Australia. Three different almond varieties were alternately planted in groups of 6 rows for cross-pollination [4] in 2006 and 2007, comprising Nonpareil (50%), Carmel (33%), and Price (17%). Plant varieties are spaced apart by 7 m and trees by 4.4 m, respectively. Nutrients were supplied to plants via drip fertigation and separated by one-hour intervals between varieties. During the 2020-2021 growing season, the fertigation rate for Nonpareil was 10% lower than that of Carmel and Price throughout the orchard.

2.2. Datasets

2.2.1. Field measurements
The field measurements and leaf sampling were conducted at the pre-harvest stage on February 1, 2021 in 13 homogeneous study plots throughout the orchard. A total of eighty fully exposed leaves of Nonpareil and Carmel were collected and measured from each study plot by different handheld instruments, including leaf $C_{ab}$, anthocyanins (Anth), flavonol content and the nitrogen balance index (NBI) using a Dualex-4 Scientific instrument (FORCE-A, Orsay, France), steady-state leaf chlorophyll fluorescence (Fl) with a FluorPen FP 110 (PSI, Brno, Czech Republic), and leaf reflectance spectra over the visible and NIR regions with a PolyPen RP 400 instrument (PSI, Brno, Czech Republic). Moreover, ten additional leaves per variety (a total of 100 leaf samples per plot) were collected for N determination in the biochemical laboratory using the Dumas Combustion method [5] with a LECO Nitrogen Analyzer (LECO Corporation, MI, USA).

2.2.2. Airborne hyperspectral imagery
An airborne campaign was carried out at solar noon under clear sky conditions on January 31, 2021. A hyperspectral line-scanning sensor (Micro-Hyperspec VNIR model, Headwall Photonics, Fitchburg, MA, USA), covering 371 bands from the visible and the near-infrared regions with an FWHM of 5.8 nm and a spectral sampling interval of 1.6 nm, was flown onboard the Cessna 172R aircraft operated by the HyperSens Laboratory, the Airborne Remote Sensing Facility of The University of Melbourne. The imagery was collected at 550 m above ground level with a spatial resolution of 40 cm. Pre-processing and calibration steps of the raw images were performed as described in Zarco-Tejada et al. [6]. The high-spatial resolution of the airborne hyperspectral imagery enabled the extraction of sunlight vegetation pixels to quantify leaf biochemistry and SIF [2]. In a two-year validation study conducted for the entire orchard, leaf N ($r^2=0.95, p<0.001$, Figure 1) was estimated with $C_{ab}$ and SIF being the most critical plant traits [3].

2.2.3. DESIS hyperspectral imagery acquired from the ISS
A 30-meter spaceborne hyperspectral scene was captured by the DESIS imaging spectrometer onboard the ISS on January 23, 2021. The imagery covers 235 spectral bands, ranging from the visible to the near-infrared regions with a 3.5 nm FWHM and a 2.55 nm sampling interval [7]. As a result of the signal noise in the blue and green spectral regions, additional cross-calibration was performed for DESIS L2A imagery using vegetation, soil and water features from airborne hyperspectral imagery by 3×3 DESIS pixel windows (Figure 2).

2.3. Plant traits retrievals and SIF quantification
The reflectance spectra of individual vegetation pixels were extracted for the calculation of structural and chlorophyll indices (e.g., NDVI, EVI, TCARI/OSAVI), as well as plant traits (e.g., $C_{ab}$, carotenoids ($C_{car}$), Anth, the de-epoxidation state of the xanthophyll-cycle pigments ($C_{x}$), dry matter ($C_{dm}$) and leaf area index (LAI)) retrieval from FluSAIL RTM [8, 9]. An artificial neural network model [10] based on a look-up-table (LUT) with random 50,000 simulations was used to retrieve the physiological plant traits.

The Fraunhofer Line Depth (FLD) principle [11] was used to calculate SIF using the $O_2$-A oxygen absorption feature from the DESIS L1C radiance imagery. Irradiance data were derived from auxiliary data collected at the nearest station on the day of DESIS overpass. The same method was applied to the airborne hyperspectral imagery to retrieve SIF and the plant traits by RTM inversions.

2.4. Statistical analysis for nitrogen estimation
The statistical methods used to assess N from airborne imagery [2, 3] were applied in this study to DESIS data. The variance inflation factor (VIF) collinearity assessment and the out-of-bag (OOB) predictor importance were also determined for DESIS. We compared the relative contributions of physiological plant traits (i.e., $C_{ab}$, $C_{car}$, Anth, ...
Cₘ, Cₘₑ, LAI) and SIF to estimate leaf N between airborne DESIS hyperspectral and airborne hyperspectral imagery throughout the almond orchard. The leaf N estimates obtained from random forest regression models using DESIS were validated with field measured leaf N and the high-resolution airborne hyperspectral-based retrievals.

3. RESULTS

The traditional structural indices (e.g., NDVI, MCARI1) calculated from DESIS hyperspectral imagery were strongly related to Cₘₑ, but yielded a weak relationship with N (Table 1). NDVI yielded $r^2=0.35$ ($p<0.05$) with leaf measured Cₘₑ, but was unable to explain N when compared either against laboratory N measurements ($r^2=0.08$, n.s.) or the airborne-based N validation map ($r^2=0.0$, n.s.). These results suggest that the variability of the canopy architecture captured at the airborne scale did not explain leaf N orchard variability. Other traditional chlorophyll indices (e.g., TCARI/OSAVI) used for N assessment did not correlate with leaf Cₘₑ nor N from DESIS data, likely due to the mixture of soil and shaded canopy components captured at such spatial resolution. Nevertheless, CTRI1 [12] exhibited a significant correlation with leaf N ($r^2=0.45$, $p<0.05$) and with the airborne-predicted N ($r^2=0.74$, $p<0.01$), outperforming other vegetation indices.

SIF quantified from DESIS showed statistically significant relationships with field measured Fᵣ ($r^2=0.52$, $p<0.01$) (data not shown), leaf Cₘₑ ($r^2=0.62$, $p<0.01$) and with leaf N ($r^2=0.56$, $p<0.01$).

Table 1. Coefficients of determination ($r^2$) for the DESIS vegetation indices against field measurements and N derived from the airborne hyperspectral imagery.

<table>
<thead>
<tr>
<th>Vegetation index</th>
<th>Leaf Cₘₑ (%)</th>
<th>Leaf PRI</th>
<th>Leaf N (%)</th>
<th>Airborne predicted N (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NDVI</td>
<td>0.35**</td>
<td>0.06</td>
<td>0.08</td>
<td>0</td>
</tr>
<tr>
<td>EVI</td>
<td>0.65***</td>
<td>0.24*</td>
<td>0.31**</td>
<td>0.20</td>
</tr>
<tr>
<td>MCARI1</td>
<td>0.69***</td>
<td>0.29*</td>
<td>0.35**</td>
<td>0.27*</td>
</tr>
<tr>
<td>SRPI</td>
<td>0.01</td>
<td>0.26</td>
<td>0.18</td>
<td>0.33**</td>
</tr>
<tr>
<td>TCARI/OSAVI</td>
<td>0.06</td>
<td>0.19</td>
<td>0.20</td>
<td>0.03</td>
</tr>
<tr>
<td>CTRI1</td>
<td>0.20</td>
<td>0.47***</td>
<td>0.45**</td>
<td>0.74***</td>
</tr>
<tr>
<td>SIF</td>
<td>0.62***</td>
<td>0.62***</td>
<td>0.56***</td>
<td>0.67***</td>
</tr>
</tbody>
</table>

*p < 0.1, **p < 0.05, ***p < 0.01

As illustrated in Table 2, Cₘₑ retrieved by RTM showed greater correlations with leaf Cₘₑ ($r^2=0.31$, $p<0.1$), leaf N ($r^2=0.63$, $p<0.01$) and airborne-predicted N ($r^2=0.80$, $p<0.01$) than chlorophyll indices. In addition to Cₘₑ, other plant traits such as Cₙ ($r^2=0.71$, $p<0.01$) were also strongly correlated with leaf N. Significant relationships were also observed between retrieved leaf pigments (i.e., Cₘₑ, Cₘ, Cₙ, Cₛ) and SIF (e.g., Cₘₑ, $r^2=0.59$, $p<0.01$). In contrast to biochemical plant traits, the structural trait LAI did not yield a significant relationship with leaf Cₘₑ nor N at DESIS scale.

Table 2. Coefficients of determination ($r^2$) among RTM-derived plant traits from DESIS and field measurements, canopy SIF and N derived from the airborne hyperspectral imagery.

<table>
<thead>
<tr>
<th>Estimated parameter</th>
<th>Leaf Cₘₑ (%)</th>
<th>Leaf N (%)</th>
<th>Airborne predicted N (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cₘₑ (µg/cm²)</td>
<td>0.31*</td>
<td>0.59***</td>
<td>0.63***</td>
</tr>
<tr>
<td>Cₘ (µg/cm²)</td>
<td>0.07</td>
<td>0.32**</td>
<td>0.17</td>
</tr>
<tr>
<td>Anth (µg/cm²)</td>
<td>0.10</td>
<td>0.01</td>
<td>0.03</td>
</tr>
<tr>
<td>Cₛ</td>
<td>0.26*</td>
<td>0.41**</td>
<td>0.71***</td>
</tr>
<tr>
<td>Cₘₑ (g/cm²)</td>
<td>0.07</td>
<td>0.12</td>
<td>0.37**</td>
</tr>
<tr>
<td>LAI</td>
<td>0.14</td>
<td>0.22</td>
<td>0.07</td>
</tr>
</tbody>
</table>

*p < 0.1, **p < 0.05, ***p < 0.01

The relative contribution of each plant trait to leaf N estimation from airborne DESIS and airborne scales appeared to be highly consistent (Figure 3). These results identified Cₘₑ, SIF, and Cₛ as the most critical spectral traits when explaining N variability, followed by the rest of the retrieved biochemical constituents and biophysical traits. Furthermore, the statistical analysis revealed that Cₘₑ and SIF were non-collinear (VIF<5) but other biochemical constituents (i.e., Cₘ, Cₘₑ and Cₘₑ) showed higher collinearity with Cₘₑ. These collinear traits were dropped from the final model to reduce redundancy. Consequently, a
model consisting of $C_{ab}$ and SIF together yielded $r^2=0.90$ ($p<0.0001$) against leaf N, and $r^2=0.75$ ($p<0.0001$) against airborne-predicted N in the almond orchard comprising different varieties, ages, and planting structures.

4. CONCLUSIONS

Results shown in this study demonstrate that RTM-derived plant traits and SIF retrieved from DESIS hyperspectral imager onboard the International Space Station yielded strong relationships with ground leaf N and with estimated N carried out from high-resolution airborne hyperspectral imagery. The most critical parameters explaining N from DESIS in this study agreed with those derived from the airborne hyperspectral imagery. Accordingly, the estimated $C_{ab}$ retrieved by RTM inversion and SIF made a greater contribution to explaining leaf N than the rest of the biochemical constituents and biophysical traits, both from DESIS and airborne hyperspectral imagery. $C_{ab}$ and SIF predicted 90% of the leaf N variability found in the almond orchard, obtaining a 75% agreement with the high-resolution airborne N estimates. The present study confirms the importance of the coupled $C_{ab}$ and SIF for leaf N assessment in tree orchards at the spaceborne scale, demonstrating the feasibility of large-scale leaf N quantification for precision agriculture purposes.

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6. REFERENCES