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Grape quality assessment in vineyards affected by iron deficiency chlorosis using narrow-band physiological remote sensing indices

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ABSTRACT

The present study investigated the use of physiological indices calculated from hyperspectral remote sensing imagery as potential indicators of wine grape quality assessment in vineyards affected by iron deficiency chlorosis. Different cv. Tempranillo/110 Richter vineyards located in northern Spain, affected and non-affected by iron chlorosis, were identified for field and airborne data collection. Airborne campaigns imaged a total of 14 study areas in both 2004 and 2005 using the AHS hyperspectral sensor, which acquired 20 spectral bands in the VIS-NIR region. Field measurements were conducted in each study site to obtain leaf and grape physiological parameters potentially linked to wine quality. Simulations carried out with the rowMCRM radiative transfer model demonstrated the feasibility of estimating leaf chlorophyll a + b (C_{ab}) content using TCARI/OSAVI from AHS spectral bands. In addition to traditional structural vegetation indices (NDVI) and successful canopy-level chlorophyll indices (TCARI/OSAVI), other innovative physiological indices sensitive to changes in carotenoid (Car) and anthocyanin ($Anth$) content in leaves were assessed from the imagery. The rowMCRM model simulations were used to evaluate canopy structural effects on these physiological indices as a function of the typical row-structured canopy variables in vineyards (LAI, crown width, row distances, C_{ab} content and soil background effects). Modeling results concluded that Car (Gitelson- $Car2$) and $Anth$ (Gitelson- $Anth$) indices were highly affected by canopy structure (C_w , V_s) and soil background (ρ_s). Field measurements of grape composition and quality were used to assess potential relationships with physiological indices sensitive to foliar pigment content (C_{ab} , Car and $Anth$). NDVI and TCARI/OSAVI indices yielded lower relationships for CIRC and IMAD must quality parameters than Car and $Anth$ physiological indices. These results suggest that the increase in carotenenes and anthocyanins due to drought, thermal damage or micronutrient deficiencies is a better indicator to detect phenolic ripening difficulties for vines affected by iron chlorosis than chlorosis detection. Therefore, the potential use of physiological remote sensing indices related to carotene and anthocyanin pigments demonstrates their importance as grape quality indicators in vineyards affected by iron chlorosis.

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1. Introduction

Iron deficiency is an important constraint in grape growing in Mediterranean climate areas. Iron deficiency decreases chlorophyll and carotenoid concentration per area in leaves (chlorosis), damaging the structure and function of the photosynthetic apparatus (Terry & Abadia, 1986). The low photosynthesis rate occurring in chlorotic plants considerably depresses the yield and vigor of vineyards (Chen & Barak, 1982; Tagliavini & Rombolà, 2001). Iron chlorosis also leads

to poor wine grape quality, reducing sugar and anthocyanin contents (Castino et al., 1987; Veliksar et al., 2005).

Grape composition plays a critical role in wine quality. In must, a large portion of the soluble solids is sugars, which determine the wine alcoholic degree. After sugars, organic acids are the most abundant solids present in grape juice. They are responsible for the tart taste and markedly influence a wine's stability and color. The aroma is created by various volatile compounds accumulated in the fruit during the ripening process and is highly dependent on environmental conditions. Phenolic compounds (anthocyanins and tannins) are present especially in skins and seeds, and determine the color and astringency of red wines. Anthocyanins are specific to red grapes and primarily responsible for wine color intensity and stability. Tannins contribute significantly to mouth-feel and color stability in red wines. Cultivar, fruit maturity, environmental conditions such as climate and soil, vineyard management practices, and the interactions among these

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Fig. 1. Examples of images of three study sites acquired in the 2004–2005 campaigns with the Airborne Hyperspectral Scanner (AHS). The images were collected at 2.5 m spatial resolution.

factors result in the composition and final potential quality of grapes (Johnson et al., 2001; Lamb et al., 2004).

The synthesis and accumulation of sugars, acids and phenolic compounds in grapes through ripening is greatly influenced by the photosynthetic capacity of the vines (Pirie & Mullins, 1980; Smart & Robinson, 1991). This capacity depends directly on vine-leaf biomass (canopy size, density and vigor) and leaf chlorophyll content (Hall et al., 2002). Research efforts in precision viticulture demonstrate the feasibility of remote sensing as a consistent method to estimate these parameters in vineyards, as potential indicators of yield (Lamb et al.,

2001) and fruit and wine composition (Johnson et al., 2001; Lamb et al., 2004; Martín et al., 2007). Precision viticulture techniques aim at pointing out homogeneous zones based on remotely sensed biophysical variable estimates, enabling inter- and within-field variability detection in vineyards and the generation of maps with a gradient of management zones potentially linked to wine quality. Previous studies showed that vineyard vigor could be successfully mapped using high spatial IKONOS satellite imagery (Johnson et al., 2003), enabling vine growth monitoring for irrigation support and canopy management through temporal relationships between the

Table 1
Hyperspectral vegetation and physiological indices used in this study.

Index	Index-ID	Equation	References
<i>Leaf area index</i>			
Normalized difference vegetation index	NDVI	$NDVI = (R_{NIR} - R_{red}) / (R_{NIR} + R_{red})$	Rouse et al. (1974)
<i>Chlorophyll estimation</i>			
Transformed C_{ab} absorption in reflectance index	TCARI	$TCARI = 3 * [(R_{700} - R_{670}) - 0.2 * (R_{700} - R_{550}) * (R_{700} / R_{670})]$	Haboudane et al. (2002)
Optimized soil-adjusted vegetation index	OSAVI	$OSAVI = (1 + 0.16) * (R_{800} - R_{670}) / (R_{800} + R_{670} + 0.16)$	Rondeaux et al. (1996)
	TCARI/OSAVI	$TCARI/OSAVI$	Haboudane et al. (2002)
	Gitelson- $Chl1$	$[R(540-560)^{-1} - R(760-800)^{-1}] * R(760-800)$	Gitelson et al. (2003, 2006)
	Gitelson- $Chl2$	$[R(690-720)^{-1} - R(760-800)^{-1}] * R(760-800)$	Gitelson et al. (2003, 2006)
<i>Carotenoid estimation</i>			
Simple ratio	Chappelle-Car	R_{760} / R_{500}	Chappelle et al. (1992)
Simple ratio	Blackburn-Car1	R_{800} / R_{470}	Blackburn (1998)
	Blackburn-Car2	$(R_{800} - R_{470}) / (R_{800} + R_{470})$	Blackburn (1998)
Structure-Intensive Pigment Index (SIPI)	SIPI	$SIPI = (R_{800} - R_{445}) / (R_{800} + R_{680})$	Peñuelas et al. (1995)
	Gitelson-Car1	$[R(510-520)^{-1} - R(540-560)^{-1}] * R(760-800)$	Gitelson et al. (2003, 2006)
	Gitelson-Car2	$[R(510-520)^{-1} - R(690-710)^{-1}] * R(760-800)$	Gitelson et al. (2003, 2006)
<i>Anthocyanin estimation</i>			
	Gamon-Anth	$R_{600-700} / R_{500-600}$	Gamon and Surfus (1999)
	Gitelson-Anth	$[R(540-560)^{-1} - R(690-710)^{-1}] * R(760-800)$	Gitelson et al. (2003, 2006)

Table 2
Hyperspectral vegetation and physiological indices calculated using AHS imagery.

Index-ID	AHS imagery adapted indices
<i>Leaf area index</i>	
NDVI	$NDVI = (R_{804} - R_{659}) / (R_{804} + R_{659})$
<i>Chlorophyll</i>	
TCARI	$TCARI = 3 * [(R_{718} - R_{689}) - 0.2 * (R_{718} - R_{571}) * (R_{718} / R_{689})]$
OSAVI	$OSAVI = (1 + 0.16) * (R_{804} - R_{689}) / (R_{804} + R_{689} + 0.16)$
TCARI/OSAVI	$TCARI / OSAVI$
Gitelson-Chl1	$[R_{571}^{-1} - R_{746}^{-1}] * R_{746}$
Gitelson-Chl2	$[R_{689}^{-1} - R_{746}^{-1}] * R_{746}$
<i>Carotenoid</i>	
Chappelle-Car	R_{746} / R_{513}
Blackburn-Car1	R_{804} / R_{484}
Blackburn-Car2	$(R_{804} - R_{484}) / (R_{804} + R_{484})$
SIPI	$SIPI = (R_{804} - R_{455}) / (R_{804} + R_{689})$
Gitelson-Car1	$[R_{484}^{-1} - R_{571}^{-1}] * R_{746}$
Gitelson-Car2	$[R_{484}^{-1} - R_{689}^{-1}] * R_{746}$
<i>Anthocyanin</i>	
Gamon-Anth	R_{659} / R_{571}
Gitelson-Anth	$[R_{571}^{-1} - R_{689}^{-1}] * R_{804}$

Normalized Difference Vegetation Index (NDVI) and the Leaf Area Index (LAI). This vegetation index and other ratios derived from field data and multispectral aerial photography were tested in recent studies to estimate canopy cover and dormant pruning weight, obtaining consistency across growing seasons (Dobrowski et al., 2002, 2003; Hall et al., 2008). These and other studies suggested broad-band multispectral remotely sensed imagery of high spatial resolution as a potential method for vineyard canopy structure characterization, enabling a successful estimation of vine canopy size, shape and row identification (Hall et al., 2003), vine mortality detection (Lagacherie et al., 2001), vineyard classification methods (Lanjari et al., 2001) and vine canopy cover estimation for irrigation management support (Montero et al., 1999). In this context, further research efforts were performed exploring high spatial resolution hyperspectral remote sensing imagery and physical methods to estimate biochemical constituents and biophysical variables as a means to assess within-field vine status and function (Zarco-Tejada et al., 2003, 2005; Martín et al., 2007).

A deeper understanding of leaf and canopy reflectance has favored the development of remote sensing applications for agriculture (Hatfield et al., 2008). Leaf biochemistry, such as the concentration

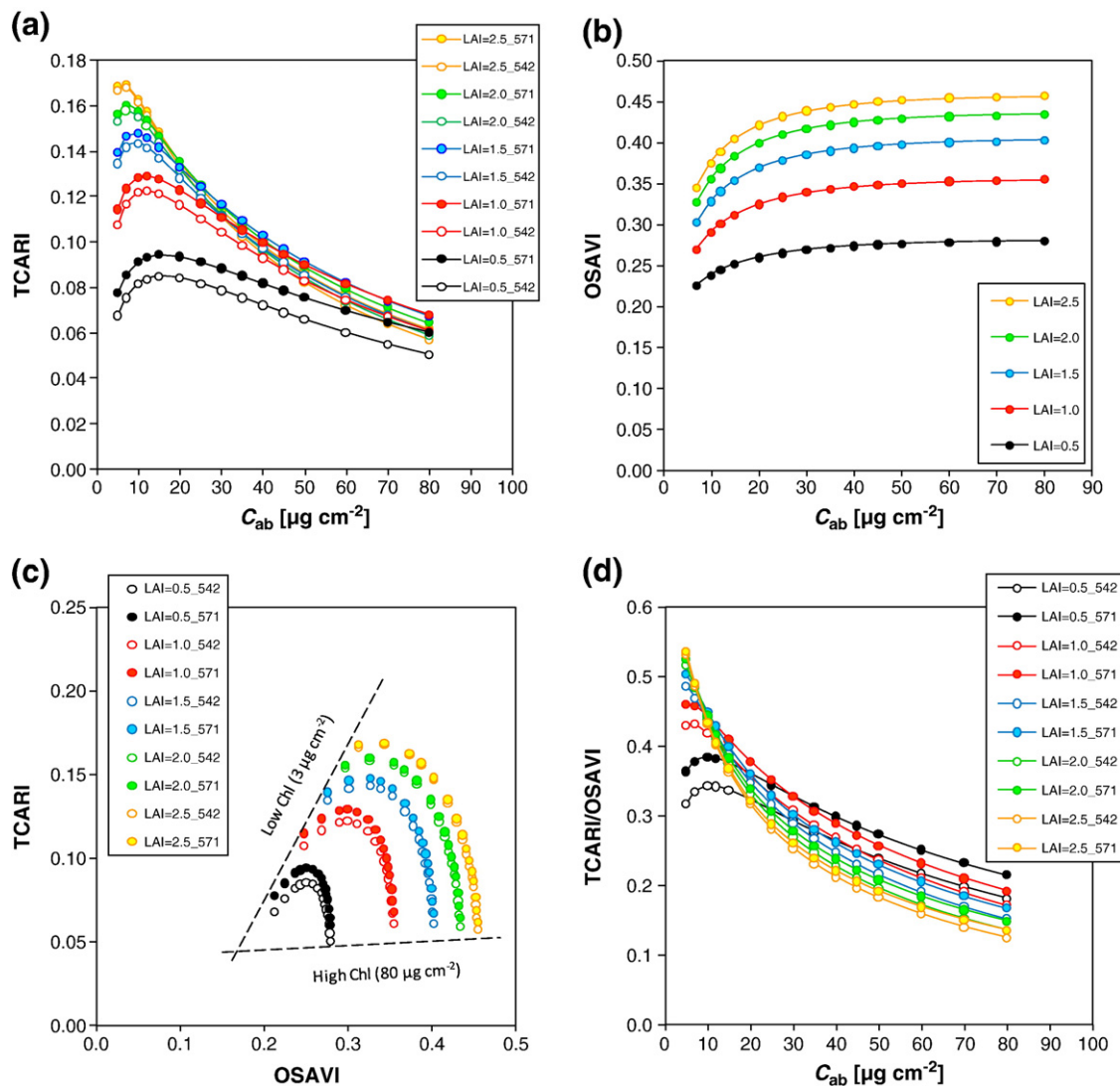


Fig. 2. Modeling simulations performed with rowMCRM to achieve effects of LAI on TCARI, OSAVI and TCARI/OSAVI sensitivity to chlorophyll variations using bands 571 nm instead of 542 nm in the TCARI index calculation.

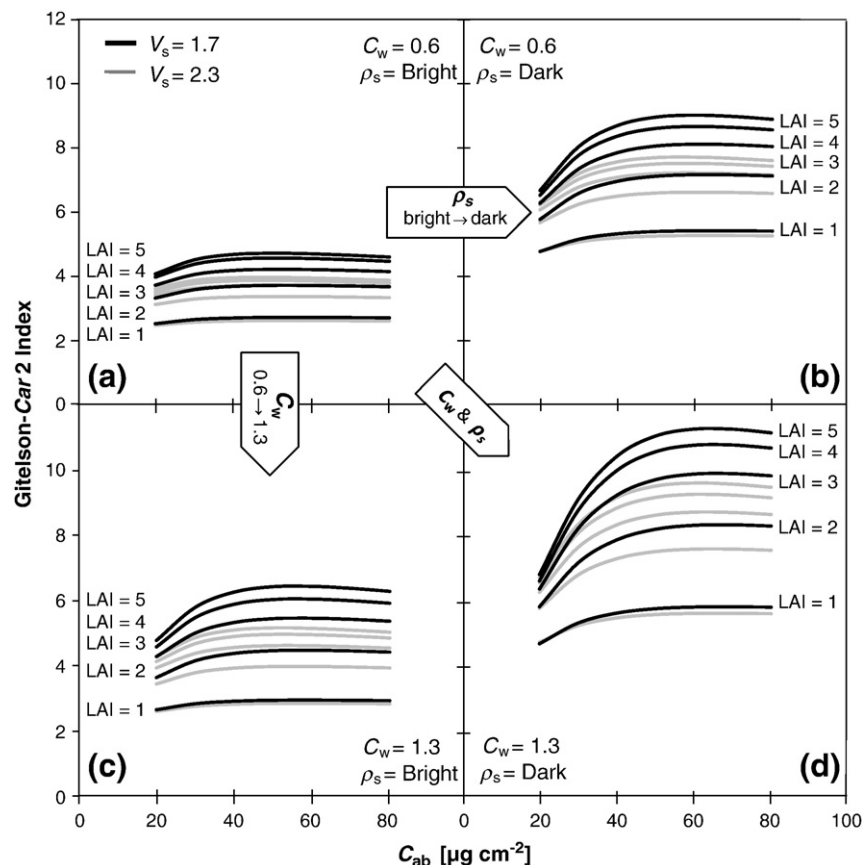


Fig. 3. Modeling simulations performed with rowMCRM to achieve the effect of soil background, LAI, vine width and visible soil strip on Gitelson-Car2 index sensitivity to chlorophyll content variation (C_{ab}).

of chlorophyll a + b (C_{ab}), water (C_w) and dry matter (C_m), is a physiological indicator of plant growth and stress status which can be estimated by empirical methods (indices) and analytical techniques (physical methods) from remote sensing data in the 400–2500 nm spectral region. Recent studies indicate that the estimation of leaf biochemical characteristics may be used as an indicator of vegetation chlorosis due to plant stress from nutritional deficiencies (Fernández-Escobar et al., 1999; Jolley & Brown, 1994; Marschner et al., 1986; Tagliavini & Rombolà, 2001; Wallace, 1991). One of the most important biochemical constituents for understanding plant physiological status is C_{ab} , which is involved in solar light energy absorption and provides the mechanism for photosynthetic reactions. Thus, C_{ab} is directly linked to photosynthetic potential and primary production, and its content in leaves is closely related to plant stress and senescence processes. Leaf C_{ab} concentration can be detected by photosynthetic pigment responses in leaf reflectance in the green peak and along the red-edge spectral region (Rock et al., 1988; Vogelmann et al., 1993; Carter, 1994; Gitelson et al., 1996).

In particular, C_{ab} estimation for chlorosis detection in vegetation has been assessed in several studies based on spectroscopy and leaf optical properties using reflectance indices, spectral and derivative indices, derivative ratios in the red-edge region (Carter & Spiering, 2002; Gitelson et al., 2003; Jacquemoud et al., 1996; le Maire et al., 2004; Sims & Gamon, 2002; Richardson et al., 2002) and new optical indices derived from high resolution hyperspectral imagery (Carter, 1994; Gitelson & Merzlyak, 1996; Vogelmann et al., 1993; Zarco-Tejada et al., 2001, 2004, 2005). In later studies, the combination of indices based on the Transformed Chlorophyll Absorption in Reflectance Index (TCARI) (Haboudane et al., 2002), the Modified

Chlorophyll Absorption in Reflectance Index (MCARI) (Daughtry et al., 2000), and the Optimized Soil-Adjusted Vegetation Index (OSAVI) (Rondeaux et al., 1996), such as TCARI/OSAVI and MCARI/OSAVI, has been demonstrated robust to soil background and leaf area index (LAI) variation in crops. These narrow-band indices, linked to radiative transfer simulations, provided useful predictive relationships for precision agriculture applications with hyperspectral imagery in continuous crop canopies (Haboudane et al., 2002) and have been adapted to open-tree canopy orchards (Zarco-Tejada et al., 2004; Meggio et al., 2008). Estimation of leaf biochemical constituents in row-structured crop canopies requires, in fact, appropriate modeling methods to account for row orientation and sun geometry, which affect the proportions of shadows, sunlit and shaded soil, and pure vegetation scene components (Zarco-Tejada et al., 2005). Differences in row-crop structural parameters, such as row height, row width, row LAI, varying soil backgrounds, and visible soil proportion, were assessed using hyperspectral imagery (Zarco-Tejada et al., 2005) to demonstrate their important effects on canopy reflectance and therefore estimation of C_{ab} . In addition, different sun viewing geometries (i.e., sun azimuth and zenith angles) have a greater influence on the optical vegetation index and the estimated leaf biochemical constituents in row-structured canopies (Meggio et al., 2008).

While most predictive relationships between leaf reflectance and pigment content have been developed for C_{ab} content estimation, providing quite robust predictions, only few models support carotenoid (Car) and anthocyanin (Anth) content estimation (Chappelle et al., 1992; Peñuelas et al., 1995; Blackburn, 1998; Gamon & Surfus, 1999; Fuentes et al., 2001; Sims & Gamon, 2002; Gitelson et al., 2001,

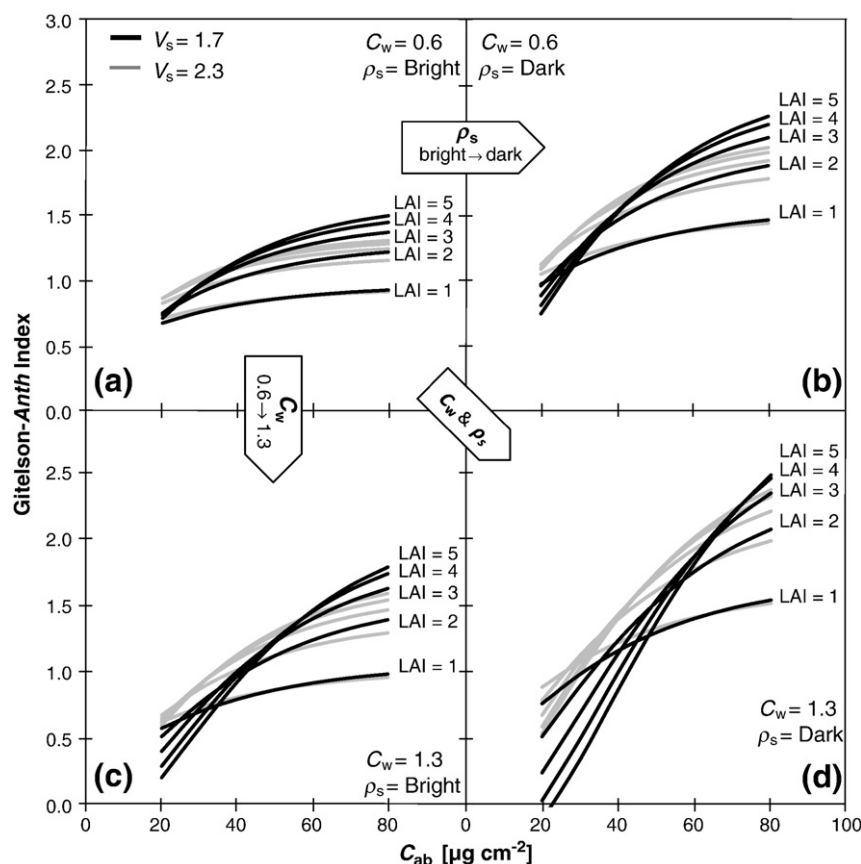


Fig. 4. Modeling simulations performed with rowMCRM to achieve the effect of soil background, LAI, vine width and visible soil strip on Gitelson-Anth index sensitivity to chlorophyll content variation (C_{ab}).

2002, 2006). *Car* and C_{ab} are the main pigments of green leaves for light harvesting, also playing a photo-protective role to prevent damage to the photosynthetic systems (Dawson et al., 1998; Gitelson et al., 2002; Merzlyak et al., 2003). *Anth*, the red pigments, protect leaves from excess light (Merzlyak & Chivkunova, 2000; Gitelson et al., 2002, 2006) and are often observed under environmental stresses such as high temperature, drought and mineral deficiencies (Chalker-Scott, 1999; Harbone, 1976). Recent studies developed physiological indices obtaining consistent relationships with *Car* (Peñuelas et al., 1995; Fuentes et al., 2001; Sims & Gamon, 2002; Gitelson et al., 2003, 2006) and *Anth* leaf pigments (Gamon & Surfus, 1999; Gitelson et al., 2006). As *Car* and *Anth* pigment concentrations are linked to photosynthetic efficiency, which affects fruit composition, physiological indices sensitive to pigment content could be used to detect spatial differences in fruit quality. In calcareous soils, iron availability can be one of the major factors modifying canopy size and foliar pigment content in leaves, which have considerable spatial variation in vineyards affected by chlorosis (Zarco-Tejada et al., 2005). Work conducted by Zarco-Tejada et al. (2005) and Martín et al. (2007) demonstrated that estimation of C_{ab} concentration at canopy level using remote sensing methods can be useful to map grape quality in vineyards affected by iron chlorosis. However, physiological indices linked to *Car* and *Anth* may yield in these areas superior results compared to other indices linked to vegetative vigor (NDVI) and C_{ab} content (TCARI/OSAVI). Therefore, research is needed on physiological condition detection using narrow-band hyperspectral remote sensing imagery. In fact, such physiological indices linked to *Car* and *Anth* have not yet been tested in vineyards for stress detection and grape quality assessment.

The objective of this investigation was to evaluate the use of physiological indices calculated from hyperspectral remote sensing imagery as potential indicators of wine grape quality assessment in vineyards affected by iron deficiency chlorosis.

2. Material and methods

2.1. Study site description

Data acquisition campaigns were conducted in July 2004 and 2005 in the western area of Ribera del Duero *Appellation d'Origine* (northern Spain). A total of seven full production vineyards, affected and non-affected by iron deficiency chlorosis, were selected for ground measurements during both years. Field data collection involved a total of 5 sub-areas of 10 m × 10 m located in each of the 7 vineyards. The study sites used for ground and airborne data collection were based on a plot network currently monitored by the local government, with specific sites selected to assure that an appropriate variability in leaf biochemistry and vine physiological conditions was found across the sites. The soils are calcareous, poor in organic matter (about 7.6 g·kg⁻¹), with a medium-weighted texture and an average pH of 8.7. Concentrations of active carbonate (up to 17.6%) and DPTA extractable Fe (1.2 to 7.6 mg·kg⁻¹) are highly heterogeneous within the area. Rainfall recorded in the study area was 358 mm in 2004 and 306 mm in 2005. All vineyards corresponded to cv. Tempranillo grafted on 110-Richter rootstock, with ages ranging between 7 and 16 years. Vine density ranged between 2200 and 4000 vines per hectare, and plants were trained to a simple or double Cordon Royat system (as described in detail in Martín et al., 2007). The vineyards

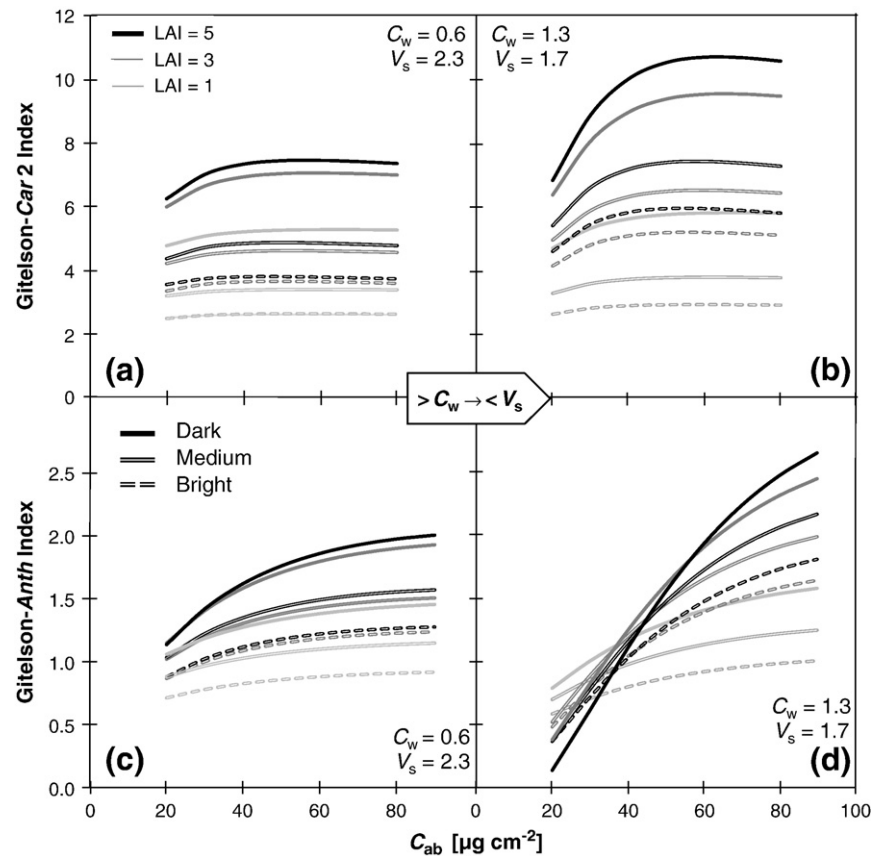


Fig. 5. Modeling simulations performed with rowMCRM to achieve the effect of soil background, LAI, vine width and visible soil strip on Gitelson-Car2 and Gitelson-Anth indices sensitivity to chlorophyll content variation (C_{ab}).

differed in canopy structure and soil background, and planting row orientation ranged from 0° to 180° N, with the row direction angle measured clockwise from north.

2.2. Physiological measurements

Field sampling campaigns were conducted in these areas concurrent with airborne overflights, comprising canopy structural parameters (grid size, number of vines within each $10\text{ m} \times 10\text{ m}$ plot, trunk height, plant height and width, and row orientation), soil chemical and structural analysis and leaf C_{ab} content determinations (Zarco-Tejada et al., 2005; Martín et al., 2007; Meggio et al., 2008). The leaf area index (LAI) and sunlit canopy cover in each study area were measured as described in Pérez (2002) and Carboneau (1995), respectively. Forty leaf samples were collected at veraison in the study zones (OIV, 1996) to determine macro- and micronutrient contents in petioles. Yield and vigor (pruning weight) of the vines were also determined.

One week before harvest, when the mean value of total soluble solid content of must in all study plots reached $228\text{ g} \cdot \text{L}^{-1}$, a total of 100 berries from each study area were collected and the must used to determine the total soluble solids content ($^\circ\text{Brix}$, total acidity (TA), tartaric and malic acid content, total polyphenols index (TPI), pH, color density and hue, in accordance with the European official methods of analysis (European Commission, 1990). In addition, a total of 100 berries collected in 2005 were sampled and their skins removed from pulp and seeds. The solid fraction was subjected to a process of extraction of polyphenolic compounds to determine the total polyphenols (Singleton & Rossi, 1965), anthocyanins (García-

Barceló, 1990), tannins (Bate-Smith, 1954), and catechin content (Swain & Hillis, 1959), while also obtaining the polymerization degree of condensed tannins (DMACH index) (Vivas et al., 1994). The extraction of polyphenolic compounds was conducted with 7.5 ml of distilled water, adding to the skins of 100 berries a total of 10 ml of acid hydroalcoholic solution (10% ethanol, 5 g L^{-1} tartaric acid). The pH was adjusted with sodium hydroxide to a value of 3.6, and 32.5 ml of distilled water were added, keeping the mixture at 35°C with slugging for 4 hours. The sample was cooled, centrifuged at 2000 g for 5 min and filtered through glass wool.

The color of musts and skin extracts was evaluated with a JASCO V-530 UV/VIS spectrophotometer. The coordinates $L^*a^*b^*$ were recorded using the D65 Illuminant as a reference (CIE, 1986). The Chromatic Index for Red Grapes ($\text{CIRG} = (180 - H^*)/(L^* + C^*)$), described by Carreño et al. (1995), was calculated.

2.3. Airborne campaigns and remote sensing physiological indices

Two years of imagery were acquired over experimental vineyards in collaboration with the Spanish Aerospace Institute (INTA) using the Airborne Hyperspectral Scanner (AHS), developed by Sensytech Inc. (currently Argon ST Inc., USA). Airborne campaigns were conducted in two consecutive years, 21 July 2004 and 21 July 2005, to acquire hyperspectral images over each vineyard. Images from both years were obtained at similar sun angles to minimize differences due to bi-directional reflectance (BRDF) effects between the years (2004, $07:45\text{--}8:22\text{ GMT}$; 2005, $09:03\text{--}09:33\text{ GMT}$). Fig. 1 shows a selection of 3 vineyard fields used in this study and imaged by the AHS airborne sensor. The hyperspectral imagery was acquired over the vineyard

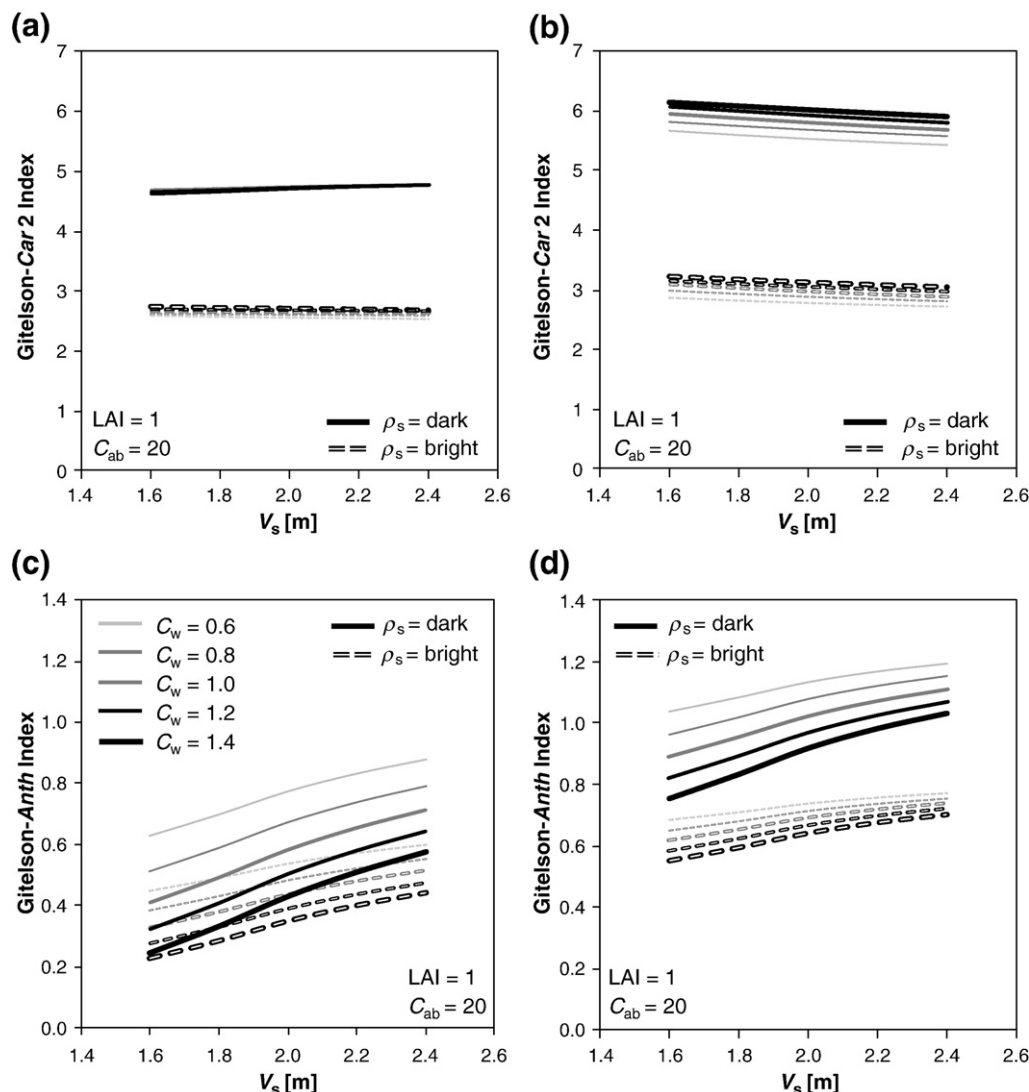


Fig. 6. Modeling simulations performed with rowMCRM to achieve the effect of the actual inter- and within-field variation of soil background (ρ_s), vine width (C_w) and chlorophyll content (C_{ab}) on Gitelson-Car2 and Gitelson-Anth indices sensitivity to visible soil variations (V_s).

fields at a flight altitude of 1000 m above ground level, obtaining images with a 90° field of view (FOV) and a 2.5 mrad instantaneous FOV (IFOV) lens, resulting in a spatial resolution of 2.5 m. The AHS sensor comprises 80 spectral bands in the 0.43–12.5 μm spectral range distributed in 4 ports (VIS/NIR, SWIR, MWIR and TIR). In this study, the VIS/NIR port was used, obtaining imagery with 20 bands over the 0.43–1.65 μm spectral range with a full-width at half-maximum (FWHM) of 30 nm.

The airborne flights were kept in the solar plane to minimize shadows, designing each overpass to image the study sites at nadir view. Images were processed by applying geometric, radiometric and atmospheric corrections. Imagery atmospheric correction was conducted using the Simple Model of Atmospheric Radiative Transfer of Sunshine (SMARTS) (Gueymard, 1995). Water vapor atmospheric content was obtained by scaling to the flight altitude the measured values at the 'Valladolid' and 'Palencia' sites as part of the AERONET network (<http://aeronet.gsfc.nasa.gov>). A full description of the calibration of the AHS bands and the radiometric and atmospheric corrections was provided in Sobrino et al. (2006). For the VIS/NIR region, soil reflectance spectra measured for each site at ground level were used to perform a flat-field correction (Ben-Dor & Levin, 2000)

after atmospheric correction, which compensated for residual effects on derived surface reflectance images estimations in the atmospheric water and oxygen absorption spectral region.

Vegetation indices were calculated from each study site to assess changes in canopy structure and foliar pigment concentration as a function of vine status and grape quality parameters. The Normalized Difference Vegetation Index (NDVI) (Rouse et al., 1974) was calculated to track changes in canopy structure given its relationship with leaf area index. The Transformed Chlorophyll Absorption in Reflectance Index (TCARI) (Haboudane et al., 2002), based on the Modified Chlorophyll Absorption in Reflectance Index (MCARI) (Daughtry et al., 2000) and normalized by the Optimized Soil-Adjusted Vegetation Index (OSAVI) (Rondeaux et al., 1996) to obtain TCARI/OSAVI, was used in this study, as it has been demonstrated to successfully minimize soil background and leaf area index variations in crops, providing predictive relationships for chlorophyll concentration estimation with hyperspectral imagery in closed crops (Haboudane et al., 2002) and open tree canopy orchards (Zarco-Tejada et al., 2004; Meggio et al., 2008).

In addition to these traditional vegetation indices, other indices more closely related to physiology were calculated through the combination of reflectance bands in the visible spectral region

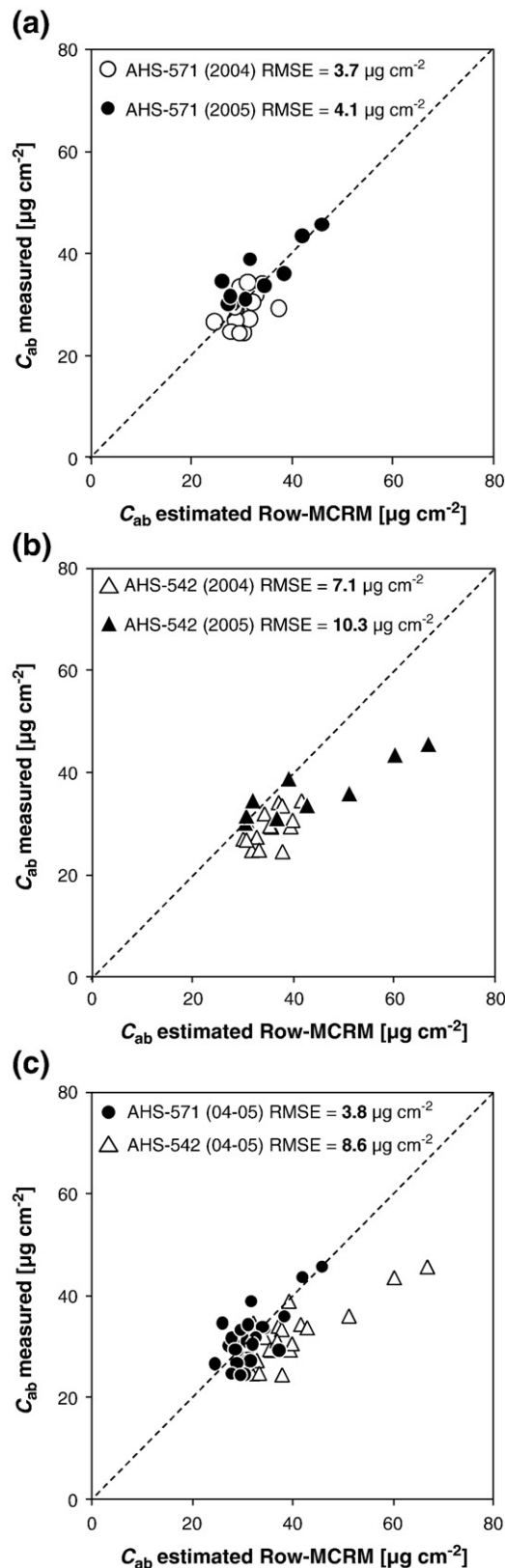


Fig. 7. Estimation of C_{ab} content at canopy level using TCARI/OSAVI through the rowMCRM model utilizing the predictive relationship proposed in Meggio et al. (2008). Differences between AHS band 571 and 542 nm for TCARI/OSAVI calculation for C_{ab} estimation.

(Gitelson et al., 2003, 2006; Chappelle et al., 1992; Blackburn, 1998; Peñuelas et al., 1995; Gamon & Surfus, 1999). These physiological indices have been proposed to track changes due to pigment content

such as chlorophyll (C_{ab}), carotenoid (Car) and anthocyanin ($Anth$). In this study, different types of indices were used as a function of sensitivity to different parameters: leaf area index (NDVI), C_{ab} content (TCARI, OSAVI, TCARI/OSAVI, Gitelson- $Chl1$, Gitelson- $Chl2$), Car content (Chappelle- Car , Blackburn- $Car1$, Blackburn- $Car2$, SIPI, Gitelson- $Car1$, Gitelson- $Car2$) and $Anth$ content (Gamon- $Anth$, Gitelson- $Anth$) (Table 1). The indices were adapted to enable calculation with AHS imagery (Table 2).

2.4. Modeling vineyard structural effects on physiological indices at the image level

Previous work demonstrates that C_{ab} content can be estimated using airborne CASI imagery through a modeling methodology to account for vineyard structure (Zarco-Tejada et al., 2003; Meggio et al., 2008). In this study, a different airborne sensor was flown over the vineyard fields, thereby requiring a simulation and validation study to assess the feasibility of the AHS bands for C_{ab} and other pigment estimation in vineyards. Previous studies, conducted with the AHS airborne sensor on PRI index changes associated with the *de-epoxidation* of the xanthophyll pigment cycle (Gamon et al., 1992), demonstrated the feasibility of the AHS bandset for TCARI/OSAVI, PRI and NDVI calculation (Zarco-Tejada et al., 2000; Suárez et al., 2008). In particular, the use of the 542 nm and 571 nm bands from the AHS airborne sensor for TCARI calculation was assessed, as the AHS bandset required a slight modification of the bands used for indices such as TCARI/OSAVI.

The MCRM model (Kuusk, 1995a,b), with additions to simulate the row vineyard structure (rowMCRM) (described in detail in Zarco-Tejada et al., 2005), was used to simulate TCARI and consequently TCARI/OSAVI sensitivity to C_{ab} with both AHS 571 nm and 542 nm bands (Fig. 2). Simulated spectra were calculated for 5 LAI values (0.5–2.5 in 0.5 steps), 15 leaf C_{ab} content levels (5–80 $\mu\text{g}\cdot\text{cm}^{-2}$ in 5 $\mu\text{g}\cdot\text{cm}^{-2}$ steps) using other rowMCRM inputs retrieved from specific data from Zarco-Tejada et al. (2003) and Meggio et al. (2008). TCARI and TCARI/OSAVI indices were calculated for each simulated spectrum using band 571 nm and 542 nm separately. The effects of plant growth (LAI) and leaf C_{ab} content on TCARI and OSAVI are illustrated in Fig. 2a, b.

As observed in Haboudane et al. (2002), LAI exerts a strong influence on the relationships between both TCARI and OSAVI and foliar pigment contents. The effects of different AHS bands used for TCARI calculation are remarkable, especially at low foliage cover ($\text{LAI} < 1$) (Fig. 2c, d). In particular, Fig. 2c shows the chlorophyll index that TCARI plotted against OSAVI for various pigment content and LAI levels, as shown in Haboudane et al. (2002), to uncouple the effects of LAI and leaf pigments separately. Both indices appear, as expected, positively correlated with LAI (low TCARI and OSAVI values correspond to low LAI values and vice versa). The different band used (542 nm, 571 nm) for TCARI calculation shows critical effects only at low LAI values ($\text{LAI} < 1$), which is important in the case of vineyard canopies. The TCARI/OSAVI index simulated with both AHS bands was then calculated, showing the trend for C_{ab} variability as a function of LAI (Fig. 2d). The simulation study enabled the selection of the AHS band 571 nm for TCARI calculation in the *scaling-up* relationship methodology proposed in Meggio et al. (2008) for C_{ab} content estimation.

The rowMCRM model was used for simulating Car and $Anth$ physiological indices in order to assess canopy structural effects on the indices as a function of the typical row-structured canopy in vineyards. Two hyperspectral indices, Gitelson- $Car2$ and Gitelson- $Anth$, sensitive to Car and $Anth$ content, respectively, were assessed as a function of vineyard structure (LAI, crown width, row distance), physiology (C_{ab} content) and soil background effects. The inputs required for canopy simulation using the rowMCRM model were the nominal ranges for leaf optical properties, canopy layer and structure,

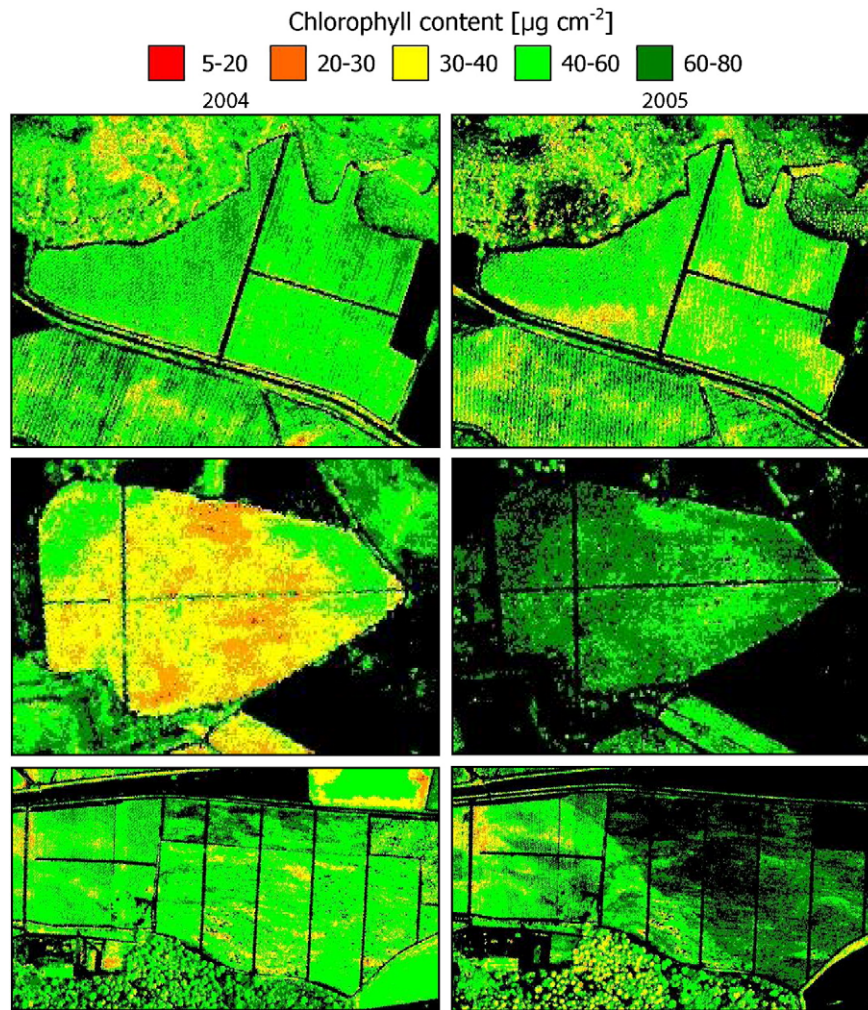


Fig. 8. Maps of spatial variation of C_{ab} content in classes of concentration ($\mu\text{g}\cdot\text{cm}^{-2}$) using the up-scaling algorithm through the TCARI/OSAVI index for the AHS 2004 (left) and 2005 (right) datasets.

and soil reflectance proposed in Zarco-Tejada et al. (2005), with the addition of row orientation effect on viewing-geometry parameters (Meggio et al., 2008).

Simulations conducted with the rowMCRM canopy reflectance model aimed at enhancing both the inter- and within-field structural and physiological variability to understand the behavior of such physiological indices in row-structured canopy conditions. In particular, different indices of the most critical rowMCRM input parameters affecting field variability were assessed, such as row-LAI, crown width (C_w), visible soil strip (V_s), leaf chlorophyll content (C_{ab}) and soil background (ρ_s) on the Gitelson-*Car2* and Gitelson-*Anth* indices. Mean vine dimensions in terms of C_w (0.6, 1.3 m) and V_s (1.7, 2.3 m) were used for all simulations.

The effects on *Car* (Gitelson-*Car2*, Fig. 3) and *Anth* (Gitelson-*Anth*, Fig. 4) indices as a function of different leaf C_{ab} content showed the effects in vine row LAI (1–5) and V_s length (1.7, 2.3 m), different ranges of C_w (0.6, 1.3 m) and ρ_s conditions (bright, dark). In particular, in both figures it can be successfully discriminated the main drivers of the inter- and within-variability affecting *Car* and *Anth* indices sensitivity to C_{ab} content variations: LAI, V_s , C_w and ρ_s . While vine row LAI and V_s variations have been reported in each single plot (a, b, c, d) presented in Figs. 3 and 4, C_w (a → b) and ρ_s (a → c) effects have been divided in order to determine the main drivers of variation. The Gitelson-*Car2* index appeared highly affected by ρ_s changes (from

bright to dark ρ_s) in all the different conditions tested, while row LAI was demonstrated to affect only at LAI > 2 (Fig. 3). Crown structural parameters C_w and V_s showed a lower effect, with ρ_s as a main driver of variation (a → d). The Gitelson-*Car2* index appeared sensitive to C_{ab} variations only in high row LAI conditions (LAI > 2), where plant structural variations also appeared to be important. The Gitelson-*Anth* index showed a similar response to the same simulations conducted for the *Car* index, confirming the higher sensitivity of C_{ab} content as a function of ρ_s (Fig. 4). In addition, the Gitelson-*Anth* index appeared more affected by C_{ab} content than the *Car* index and less affected by LAI and ρ_s at low C_{ab} content, increasing with a further change in C_{ab} .

Therefore, the effects of soil background on the physiological indices were assessed, showing the large effects due to ρ_s on the *Car* (Gitelson-*Car2*) and *Anth* (Gitelson-*Anth*) indices as a function of different C_{ab} levels (Fig. 5). A range of 3 different ρ_s spectra from bright to dark collected in the field over the same area were tested in simulations for different row LAI (1–5) in 2 opposite structural conditions in terms of V_s and C_w . These simulations assessed the high effect of ρ_s on both physiological indices as a function of C_{ab} content, with a greater effect represented by a higher vine crown dimension (C_w = 1.3 m) and therefore a lower visible soil strip (V_s = 1.7 m) conditions.

The wide range simulation results presented in Figs. 3–5 underlined that *Car* (Gitelson-*Car2*) and *Anth* (Gitelson-*Anth*) indices are

Table 3Coefficients of determination (r^2) found for linear relationships between hyperspectral indices and grapevine structural parameters in 2004 and 2005 ($p < 0.05$).

Index-ID	Reference	Grapevine structural and management parameters							
		ESA		LAI		Yield		Pruning weight	
		04	05	04	05	04	05	04	05
<i>Leaf area index</i>									
NDVI	Rouse et al. (1974)		0.62+						0.44+
<i>Chlorophyll</i>									
TCARI/OSAVI	Haboudane et al. (2002)								0.70+
Gitelson-Chl1	Gitelson et al. (2003, 2006)		0.60+						
Gitelson-Chl2	Gitelson et al. (2003, 2006)								0.51+
<i>Carotenoid</i>									
Chappelle-Car	Chappelle et al. (1992)			0.47–					
Blackburn-Car1	Blackburn (1998)			0.46–					
Blackburn-Car2	Blackburn (1998)		0.41+	0.48–					
SIPI	Peñuelas et al. (1995)			0.41–					
Gitelson-Car1	Gitelson et al. (2003, 2006)						0.55–		
Gitelson-Car2	Gitelson et al. (2003, 2006)						0.58–		
<i>Anthocyanin</i>									
Gamon-Anth	Gamon and Surfus (1999)								0.65–
Gitelson-Anth	Gitelson et al. (2003, 2006)					0.44–	0.55+		

ESA = Exposed Canopy Surface Area; LAI = Leaf Area Index.

affected by structure (C_w , V_s) and soil background (ρ_s). New simulations were conducted with rowMCRM to assess the expected real field variability, considering the actual range for C_{ab} ($20\text{--}40\ \mu\text{g}\cdot\text{cm}^{-2}$), C_w ($0.6\text{--}1.4\text{ m}$ in 0.2 m steps), V_s ($1.6\text{--}2.4\text{ m}$ in 0.2 m steps) and ρ_s (bright, dark). The effect on *Car* and *Anth* indices as a function of V_s dimensions was studied for different C_w , ρ_s and C_{ab} conditions to assess the expected real field variability (Fig. 6). Simulations conducted revealed the sensitivity of Gitelson-Car2 (Fig. 6a,b) and Gitelson-Anth (Fig. 6c,d) as a function of V_s to ρ_s in both low ($20\ \mu\text{g}\cdot\text{cm}^{-2}$) to medium ($40\ \mu\text{g}\cdot\text{cm}^{-2}$) C_{ab} content. Fig. 6c, d underlines that the higher sensitivity of the *Anth* index to C_{ab} variations is due mainly to ρ_s . The modeling study demonstrated the influence of structure on physiological indices used for *Car* and *Anth* assessment. *Scaling-up* relationships for C_{ab} estimation and results on

the assessment of the physiological indices used for grape quality estimation are described in the following section.

3. Results

3.1. Chlorosis detection using rowMCRM modeling with the AHS airborne imager

Previous field campaigns conducted in 2003 over the same study sites using the rowMCRM simulation methodology demonstrated the capability of the CASI hyperspectral airborne sensor for C_{ab} estimation in row-structured canopies (Zarco-Tejada et al., 2005; Meggio et al., 2008). Results obtained following the same methodology on the AHS sensor bandset are presented. The predictive relationship developed

Table 4Coefficients of determination (r^2) found for linear relationships between hyperspectral indices and must composition parameters in 2004 and 2005 ($p < 0.05$).

Index-ID	Reference	Must composition parameters													
		W100		°BRIX		TA		pH		TPI		Tartaric		Malic	
		04	05	04	05	04	05	04	05	04	05	04	05	04	05
<i>Leaf area index</i>															
NDVI	Rouse et al. (1974)			0.50+						0.62+					
<i>Chlorophyll</i>															
TCARI/OSAVI	Haboudane et al. (2002)			0.50–								0.53–			
Gitelson-Chl1	Gitelson et al. (2003, 2006)		0.65+	0.52+						0.62+					
Gitelson-Chl2	Gitelson et al. (2003, 2006)			0.53+						0.60+					
<i>Carotenoid</i>															
Chappelle-Car	Chappelle et al. (1992)	0.49+	0.49+	0.47–						0.59+				0.47–	
Blackburn-Car1	Blackburn (1998)	0.49+	0.50+	0.48+						0.57+				0.44–	
Blackburn-Car2	Blackburn (1998)	0.52+	0.48+	0.43+		0.44+				0.57+				0.47–	
SIPI	Peñuelas et al. (1995)		0.63+	0.50+						0.62+					
Gitelson-Car1	Gitelson et al. (2003, 2006)					0.51+		0.63–				0.45+		0.81–	
Gitelson-Car2	Gitelson et al. (2003, 2006)							0.65–						0.78–	
<i>Anthocyanin</i>															
Gamon-Anth	Gamon and Surfus (1999)			0.48–				0.70–	0.53–					0.67–	
Gitelson-Anth	Gitelson et al. (2003, 2006)			0.46–				0.66–						0.78–	

W100: 100 berries weight; TA: Total Acidity; TPI: Total Polyphenol Index; Tartaric Tartaric acid content; Malic Malic acid content.

Table 5Coefficients of determination (r^2) found for linear relationships between hyperspectral indices and color intensity (CI), hue and CIELAB coordinates for must ($p < 0.05$).

Index-ID	Reference	Must color parameters (2004)						
		<i>L</i>	<i>a</i> *	<i>b</i> *	<i>c</i> *	H	CI	Hue
<i>Leaf area index</i>								
NDVI	Rouse et al. (1974)	0.69—	0.68+		0.72+	0.73—		
<i>Chlorophyll</i>								
TCARI/OSAVI	Haboudane et al. (2002)	0.68—						
Gitelson- <i>Chl1</i>	Gitelson et al. (2003, 2006)	0.74—	0.74+		0.81+	0.70—		
Gitelson- <i>Chl2</i>	Gitelson et al. (2003, 2006)	0.71—	0.71+		0.75+	0.74—		
<i>Carotenoid</i>								
Chappelle-Car	Chappelle et al. (1992)	0.68—	0.7+		0.80+	0.57—	0.82+	0.74—
Blackburn-Car1	Blackburn (1998)	0.64—	0.69+		0.80+	0.55—	0.72+	0.72—
Blackburn-Car2	Blackburn (1998)	0.72—	0.64+		0.74+	0.50—	0.67+	0.68—
SIPI	Peñuelas et al. (1995)		0.71+		0.78+	0.69—		0.68—
Gitelson-Car1	Gitelson et al. (2003, 2006)							
Gitelson-Car2	Gitelson et al. (2003, 2006)							
<i>Anthocyanin</i>								
Gamon-Anth	Gamon and Surfus (1999)	0.58+	0.58—	0.45+	0.59—	0.76+		
Gitelson-Anth	Gitelson et al. (2003, 2006)					0.64+	0.70—	0.55+

by Meggio et al. (2008) with the rowMCRM model between C_{ab} and TCARI/OSAVI were adapted for the AHS bandset.

Simulations conducted through the rowMCRM model assessed the effect of using AHS bands 571 nm and 542 nm for TCARI calculation. Leaf C_{ab} content was estimated using the predictive scaling relationship through AHS TCARI/OSAVI calculated with both bands separately, assessing the bandset effects on the root mean square error (RMSE) for C_{ab} estimation (Fig. 7). TCARI/OSAVI calculated with band 542 nm obtained an RMSE of 7.1 and 10.3 $\mu\text{g} \cdot \text{cm}^{-2}$ for the AHS 2004 and 2005 datasets confirming, in any case, the reliability of the scaling relationship (Fig. 7a). C_{ab} estimation through AHS band 571 nm for TCARI/OSAVI calculation yielded better results from both the 2004 and 2005 datasets, with an RMSE of 3.7 and 4.1 $\mu\text{g} \cdot \text{cm}^{-2}$, respectively (Fig. 7b). Results obtained suggested, therefore, the use of AHS band

571 nm for TCARI/OSAVI calculation for C_{ab} estimation, yielding an overall RMSE of 3.8 $\mu\text{g} \cdot \text{cm}^{-2}$ considering the two study years together, instead of band 542 nm, which yielded an overall RMSE of 8.6 $\mu\text{g} \cdot \text{cm}^{-2}$ (Fig. 7c).

These results demonstrate the feasibility of estimating C_{ab} with the AHS airborne sensor, an instrument with 30 nm FWHM bands, through the modeling methodology presented. The scaling algorithm for TCARI/OSAVI to estimate C_{ab} content proposed in Meggio et al. (2008), considering both before and after noon viewing geometries and differences in vine row orientation, was applied to the AHS images to obtain maps of leaf C_{ab} content for five levels, ranging from 5 to 80 $\mu\text{g} \cdot \text{cm}^{-2}$ (Fig. 8). These C_{ab} maps show the within- and inter-field variability in vineyards affected by iron deficiency chlorosis, displaying different chlorosis levels for both years.

Table 6Coefficients of determination (r^2) found for linear relationships between hyperspectral indices and must quality indices ($p < 0.05$).

Index-ID	Reference	Must quality indices							
		QI-1		QI-2		IMAD		CIRG	
		04	05	04	05	04	05	04	05
<i>Leaf area index</i>									
NDVI	Rouse et al. (1974)			0.63—					
<i>Chlorophyll</i>									
TCARI/OSAVI	Haboudane et al. (2002)								
Gitelson- <i>Chl1</i>	Gitelson et al. (2003, 2006)		0.57—	0.67+					
Gitelson- <i>Chl2</i>	Gitelson et al. (2003, 2006)			0.62+					
<i>Carotenoid</i>									
Chappelle-Car	Chappelle et al. (1992)		0.53—	0.68+			0.44—		0.50—
Blackburn-Car1	Blackburn (1998)		0.54—	0.70+			0.44—		0.48—
Blackburn-Car2	Blackburn (1998)		0.55—	0.68+			0.48—		0.51—
SIPI	Peñuelas et al. (1995)		0.58—	0.65+					
Gitelson-Car1	Gitelson et al. (2003, 2006)	0.48—				0.59—	0.42—	0.45—	0.92—
Gitelson-Car2	Gitelson et al. (2003, 2006)					0.71—	0.42—	0.68—	0.93—
<i>Anthocyanin</i>									
Gamon-Anth	Gamon and Surfus (1999)			0.54—				0.43—	0.64—
Gitelson-Anth	Gitelson et al. (2003, 2006)						0.48—	0.56—	0.94—

%vol: probable alcoholic degree.

QI-1: %vol/100 berries weight; QI-2: $(\text{TPI}/40)^2 \times (\% \text{vol} - 10)$; CIRG: $(180 - H^*)/(L^* + C^*)$; IMAD: °Brix/TA.

Table 7Coefficients of determination (r^2) found for linear relationships between hyperspectral indices and berry skins composition ($p < 0.05$).

Index-ID	Reference	Skin extracts components and quality parameters (2005)						
		W100 Skins	DMACH Index	Tannins	Catechins	AT	PT	CIRG
<i>Leaf area index</i>								
NDVI	Rouse et al. (1974)		0.72—					
<i>Chlorophyll</i>								
TCARI/OSAVI	Haboudane et al. (2002)							
Gitelson-Chl1	Gitelson et al. (2003, 2006)	0.53+	0.50—		0.51—		0.47—	
Gitelson-Chl2	Gitelson et al. (2003, 2006)		0.67—					
<i>Carotenoid</i>								
Chappelle-Car	Chappelle et al. (1992)	0.50+			0.57—	0.66—	0.63—	0.70—
Blackburn-Car1	Blackburn (1998)	0.52+			0.58—	0.61—	0.62—	0.70—
Blackburn-Car2	Blackburn (1998)	0.48+			0.55—	0.67—	0.62—	0.66—
SIPI	Peñuelas et al. (1995)	0.51+	0.60—		0.50—		0.58—	
Gitelson-Car1	Gitelson et al. (2003, 2006)			0.44—		0.75—	0.48—	0.68—
Gitelson-Car2	Gitelson et al. (2003, 2006)			0.48—		0.75—	0.47—	0.68—
<i>Anthocyanin</i>								
Gamon-Anth	Gamon and Surfus (1999)			0.59—		0.44—		0.45—
Gitelson-Anth	Gitelson et al. (2003, 2006)			0.52—		0.71—	0.48—	0.68—

W100 skins: 100 skins weight; Tannins: Total Tannins content; AT: Total Anths content; PT: Total Polyphenols content.

3.2. Relationships between hyperspectral indices and physiological data for grape quality assessment

Physiological measurements conducted for two years on grapes sampled one week before harvest were used to assess the relationship with hyperspectral indices sensitive to pigment content (C_{ab} , Car , and $Anth$). Coefficients of determination (r^2) found for linear regression relationships between hyperspectral indices calculated from AHS imagery and grape and must quality parameters were obtained for four types of indices: leaf area index and structural indices (i), C_{ab} (ii), Car (iii) and $Anth$ content (iv). Coefficients of determination values

resulting from linear regression are presented in Tables 3–7, along with the direct/inverse relationship (+ or –). The relationships with $r^2 < 0.4$ and $p > 0.05$ are not included in the tables (relationships with $r^2 > 0.6$ are underlined in bold).

Plant structural and management parameters (LAI, exposed canopy surface area (ESA), yield and pruning weight) were related to vegetation indices (Table 3). Linear relationships were obtained between hyperspectral indices generally considered sensitive to vegetative vigor and grapevine structural parameters. In particular, TCARI/OSAVI and NDVI vegetation indices yielded coefficients of determination with pruning weight ($r^2 = 0.7$) and exposed canopy surface ($r^2 = 0.62$), respectively. Mean mineral concentrations in petioles were similar or higher than reference values for Ribera del Duero Appellation d'Origine (González and Martín, 2006). However, there were wide ranges of variation within samples for Mn, Cu and Zn (data not shown), including some study subzones with deficiencies in these micronutrients. Significant correlations between Zn foliar content and yield and vigor of vineyards in the same study area have been reported previously (Martín et al., 2008).

Relationships with must composition parameters ($^{\circ}$ BRIX, pH, total acidity (TA), tartaric and malic acid content) are shown in Table 4, displaying those obtained with the hyperspectral indices tested in this study. While the traditional NDVI index yielded relationships with $^{\circ}$ BRIX ($r^2 = 0.5$) and the Total Polyphenol Index – TPI ($r^2 = 0.62$), other physiological indices, Gamon-Anth and Gitelson-Car1, yielded significant linear relationships with pH ($r^2 = 0.7$) and malic acid content ($r^2 = 0.81$), respectively. Coefficients of determination obtained for chromatic characteristics of must (Table 5) show that physiological indices appeared more suitable for color intensity (CI) and hue estimation than traditional vegetation indices. Linear relationships performed resulted in high coefficients of determination with CI ($r^2 = 0.82, 0.72, 0.70$) and hue ($r^2 = 0.74, 0.72, 0.55$) for the Chappelle-Car, Blackburn-Car1 and Gitelson-Anth physiological indices, respectively. Different must quality indices, such as IMAD ($^{\circ}$ Brix/TA) and the Chromatic Index for Red Grapes (CIRG) (Table 6), show consistent results obtained for both seasons, 2004 and 2005. In particular, unlike the results achieved with traditional (NDVI) and more innovative (TCARI/OSAVI) indices sensitive to vegetative vigor and C_{ab} content, physiological indices sensitive to Car and $Anth$ content appeared more suitable for grape quality assessment in the vineyards studied. Specifically, the inverse linear relationships

Table 8

Hyperspectral indices with the highest coefficients of determination for each grapevine quality parameter.

Must composition		Must quality	
W100	Gitelson-Chl1 SIPI	QI-1	SIPI Gitelson-Chl1
$^{\circ}$ BRIX	Gitelson-Chl1-2	QI-2	Blackburn-Car1-2
pH	Gamon-Anth	IMAD	Chappelle-Car
TPI	Gitelson-Anth		Gitelson-Car1-2
Malic	Gitelson-Chl1 SIPI	CIRG	Gitelson-Anth
	Gitelson-Car1-2		Gitelson-Car1-2
	Gitelson-Anth		
Must color		Skin extracts components and quality	
L	Gitelson-Chl1	W100 skins	Gitelson-Chl1
	Blackburn-Car2		Blackburn-Car1
a^*	Gitelson-Chl1-2 SIPI	DMACH index	Gitelson-Chl2
c^*	Gitelson-Chl1	Tannins	Gamon-Anth
	Chappelle-Car		
	Blackburn-Car1		
H	Gamon-Anth	Catechins	Chappelle-Car
	Gitelson-Chl2		Blackburn-Car1
CI	Chappelle-Car	AT	Gitelson-Car1-2
	Blackburn-Car1		Gitelson-Anth
Hue	Chappelle-Car	PT	Chappelle-Car
	Blackburn-Car1	CIRG	Blackburn-Car1-2
			Chappelle-Car
			Blackburn-Car1
			Gitelson-Car1-2
			Gitelson-Anth

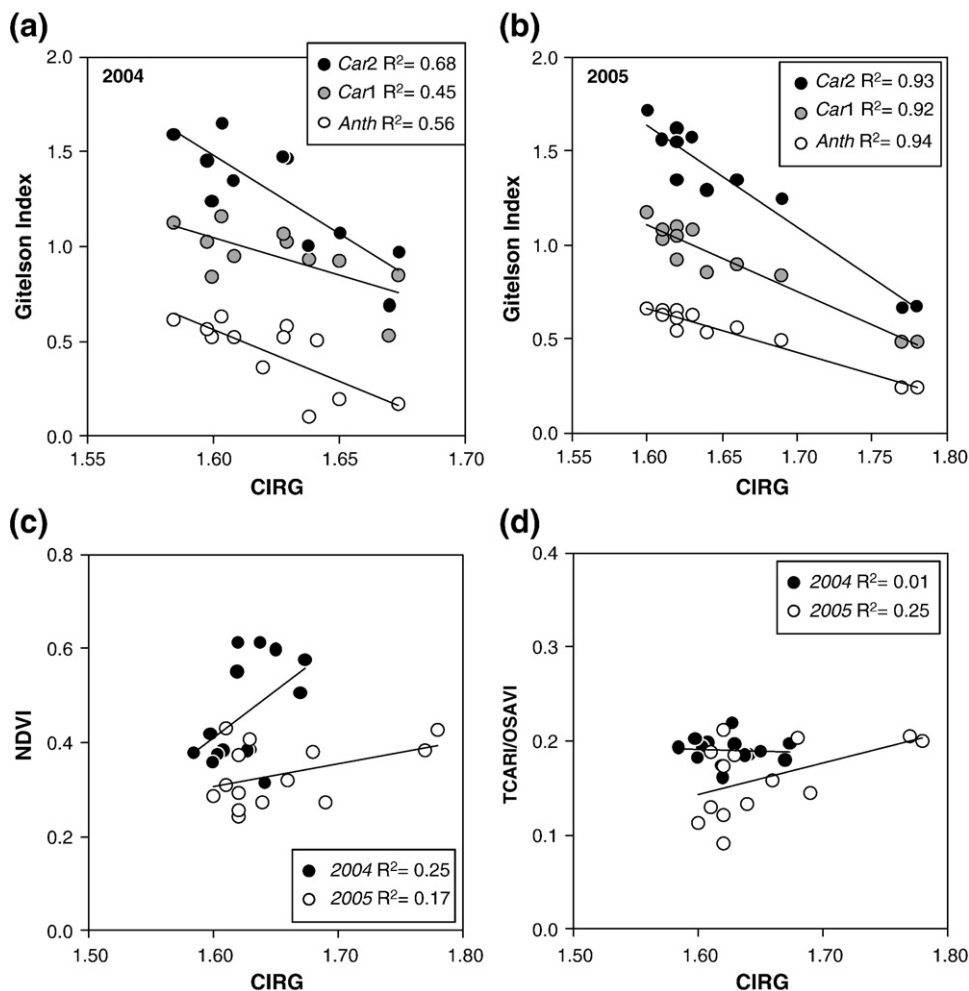


Fig. 9. Relationships between the must Chromatic Index for Red Grapes (CIRG) and physiological indices sensitive to *Car* and *Anth* pigment content (a,b) and traditional indices sensitive to C_{ab} content (c,d).

obtained between CIRG and the physiological indices yielded consistent coefficients of determination, $r^2 = 0.68$ (2004) and $r^2 = 0.93$ (2005) for Gitelson-*Car2*, and $r^2 = 0.56$ (2004) and $r^2 = 0.94$ (2005) for the Gitelson-*Anth* index. The IMAD quality index yielded interesting linear relationships with the Gitelson-*Car2* physiological index, obtaining $r^2 = 0.71$ and $r^2 = 0.42$ for the 2004 and 2005 season datasets, respectively. The Gitelson-*Car2* and Gitelson-*Anth* indices were inversely correlated with B foliar levels in 2004 (Pearson's $r = -0.89$, $p < 0.01$) and with K, Cu, Zn and B foliar levels in 2005 (r values from -0.61 to -0.88 , $p < 0.05$).

Finally, results obtained for berry skin composition parameters in the 2005 field campaign (100 skins weight, DMACH index, total tannin index, total *Anth* (AT), total polyphenols (PT) and CIRG) (Table 7) show that, while the traditional NDVI vegetation index appeared well correlated ($r^2 = 0.72$) only with the DMACH index, hyperspectral indices sensitive to physiological properties were shown to be more suitable for berry skin composition estimation. In particular, indices sensitive to *Car* and *Anth* content appeared well correlated (negatively) with AT, PT and CIRG physiological parameters. Consistent relationships have been obtained between Gitelson indices and berry skin AT content: $r^2 = 0.75$ (Gitelson-*Car*) and $r^2 = 0.71$ (Gitelson-*Anth*). The berry skin PT appeared well correlated with the Chappelle-*Car* and Blackburn-*Car* indices, yielding coefficients of determination of $r^2 = 0.63$ and 0.62 , respectively. The skin

CIRG index for skin extracts was inversely well correlated with the Chappelle-*Car* ($r^2 = 0.7$) and Gitelson-*Anth* ($r^2 = 0.68$) physiological indices.

Positive Pearson's coefficients between total anthocyanin concentrations in skin extracts and K ($r = 0.54$, $p < 0.05$), Cu ($r = 0.69$, $p < 0.01$), Zn ($r = 0.76$, $p < 0.001$) and B ($r = 0.52$, $p < 0.05$) content in petioles were observed. Total polyphenols also were correlated with Cu and B levels ($r = 0.75$ and $r = 0.72$, respectively, $p < 0.01$). The best relationships were obtained for Zn-Gamon-*Anth* ($r = -0.91$) and B-Blackburn-*Car2* ($r = -0.86$).

In summary, these results suggest that traditional vegetation indices sensitive to leaf area index (NDVI) and newer indices related to C_{ab} content (TCARI/OSAVI) yielded lower relationships than physiological indices sensitive to carotenoid *Car* and anthocyanin *Anth* content. In particular, the comparison between physiological indices potentially useful for estimating grape quality showed relationships for the two years between must and skin composition parameters and *Car* and *Anth* indices (Gitelson-*Car1*, *Car2* and Gitelson-*Anth* indices, Chappelle-*Car*, Blackburn-*Car1*, *Car2*) (Table 8). The best relationships were obtained for wine grape quality parameters, as presented in Figs. 9 to 13. The relationships between the chromatic index (CIRG) and *Car*, *Anth* indices, TCARI/OSAVI and NDVI (Fig. 9) demonstrate the capability of physiological indices for grape quality assessment. While the Gitelson-*Car2* and Gitelson-*Anth*

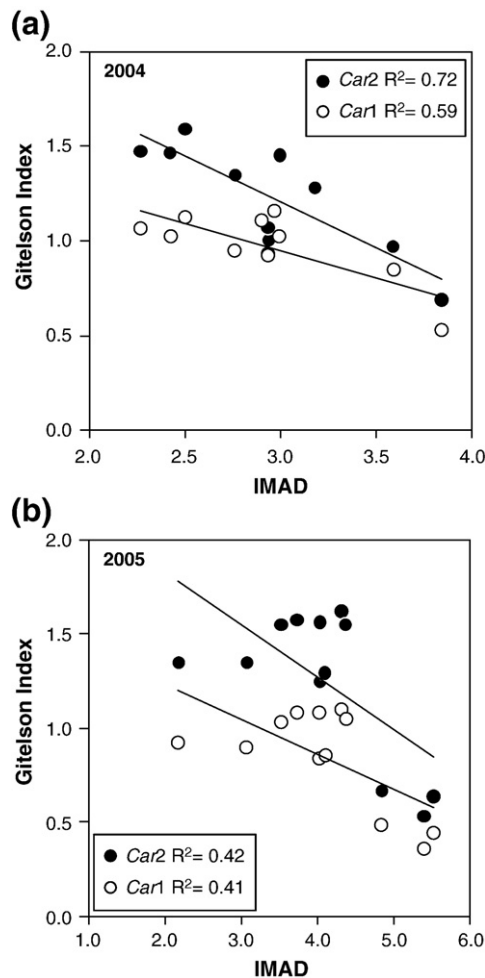


Fig. 10. Relationships between the IMAD (°vol/total acidity) Index and physiological indices sensitive to Car and Anth pigment content.

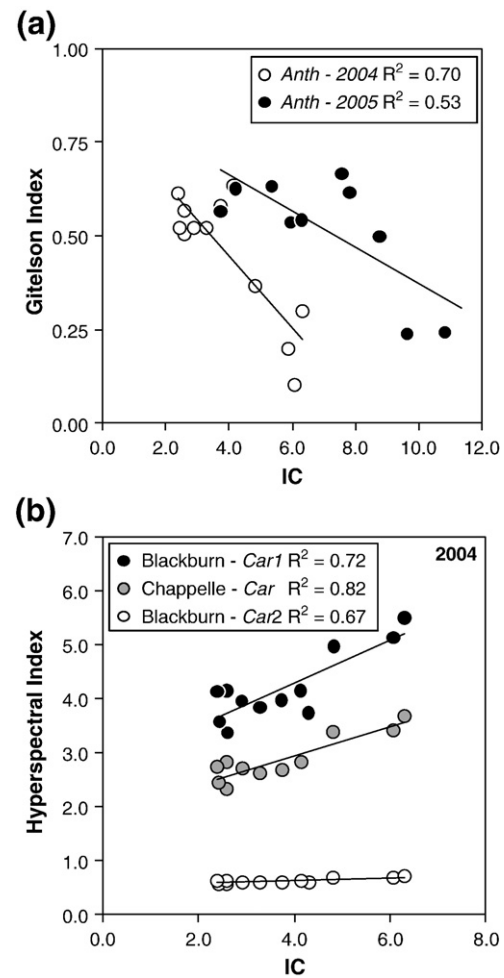


Fig. 11. Relationships between must color intensity (CI) and physiological indices sensitive to Car and Anth pigment content.

indices yielded coefficients of determination ranging between $r^2 \sim 0.5$ – 0.6 and $r^2 \sim 0.9$ with CIRG for the 2004 and 2005 seasons, respectively, NDVI and TCARI/OSAVI showed lower relationships ($r^2 < 0.25$).

The relationships between the IMAD index and Gitelson-Car indices (Fig. 10) yielded coefficients of determination ranging between $r^2 \sim 0.6$ – 0.7 and $r^2 \sim 0.4$ for the 2004 and 2005 seasons, respectively. Must color intensity (CI) obtained consistent relationships with the Gitelson-Anth index for both the 2004 ($r^2 = 0.7$) and 2005 ($r^2 = 0.53$) growing seasons (Fig. 11). Consistent relationships were obtained between berry skin total polyphenols (PT) collected in the 2005 season and physiological indices sensitive to Car content, with a coefficient of determination of $r^2 \sim 0.6$ for the Chappelle-Car, Blackburn-Car1-2 and SIPI indices (Fig. 12). The berry skin Chromatic Index for Red Grapes (CIRG) resulted in consistent relationships with both Car and Anth indices, yielding a coefficient of determination of around $r^2 = 0.68$ – 0.7 (Fig. 13).

The quantitative results obtained in Figs. 9 and 10 enabled the calculation of spatial maps of grape quality for CIRG (Fig. 14) and IMAD (Fig. 15) indices. The maps were obtained through Gitelson-Car2 index linear relationships with CIRG(2004) [$y = -8.252(x) + 14.685$], CIRG(2005) [$y = -5.4395(x) + 10.344$], IMAD(2004) [$y = -0.4898(x) + 2.6772$] and IMAD(2005) [$y = -0.2788(x) + 2.3851$] quality indices for three vineyard study sites, showing both the inter- and within-field variability between the different vineyards for two consecutive years.

4. Discussion

Simulations conducted with the rowMCRM model demonstrated the feasibility of estimating C_{ab} content using TCARI/OSAVI from AHS 30 nm FWHM spectral bands. Results underlined the strong effect of within- and between-field variability on the TCARI/OSAVI hyperspectral index when calculated with band 571 nm instead of band 541 nm for the TCARI index. Leaf C_{ab} content estimated from AHS TCARI/OSAVI calculated with band 571 nm through the scaling algorithm proposed by Meggio et al. (2008) provided the best results, yielding an RMSE of 3.7 and 4.1 $\mu\text{g}\cdot\text{cm}^{-2}$ for the 2004 and 2005 datasets, respectively, and resulting in an overall RMSE of 3.8 $\mu\text{g}\cdot\text{cm}^{-2}$ (RMSE = 8.6 $\mu\text{g}\cdot\text{cm}^{-2}$ if using band 542 nm). Such consistent results enabled the calculation of C_{ab} content distribution maps in 5 levels of chlorophyll concentration (5–80 $\mu\text{g}\cdot\text{cm}^{-2}$).

The rowMCRM model simulations were used to assess canopy structural effects on physiological indices sensitive to Car and Anth content as a function of the typical row-structured canopy variables in vineyards (LAI, crown width, row distance, C_{ab} content and soil background effects) and then consider the expected real field variability and actual range parameters collected in the field sites. Modeling results concluded that Car (Gitelson-Car2) and Anth (Gitelson-Anth) indices were highly affected by canopy structure (C_w , V_s) and soil background (ρ_s).

Coefficients of determination resulting from the linear regression between field parameters and physiological indices (in both years of

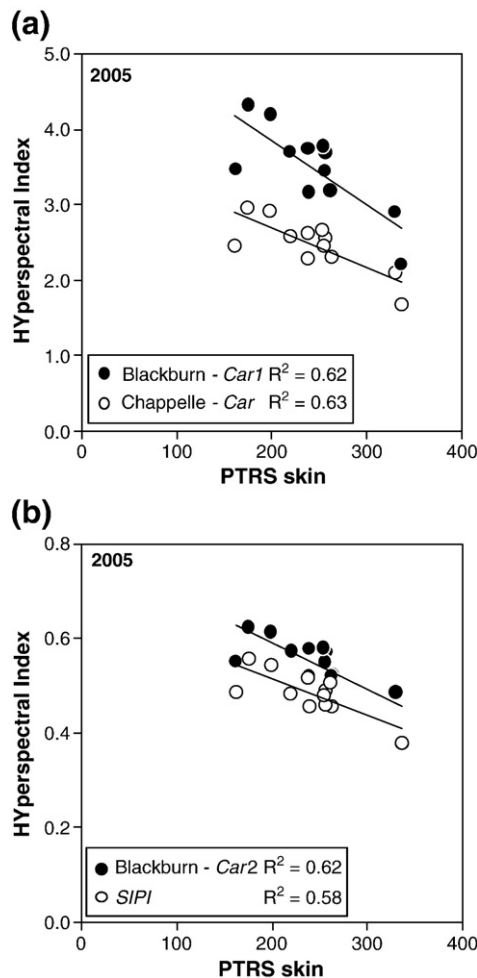


Fig. 12. Relationships between berry skin total polyphenols content and physiological indices sensitive to Car and Anth pigment content.

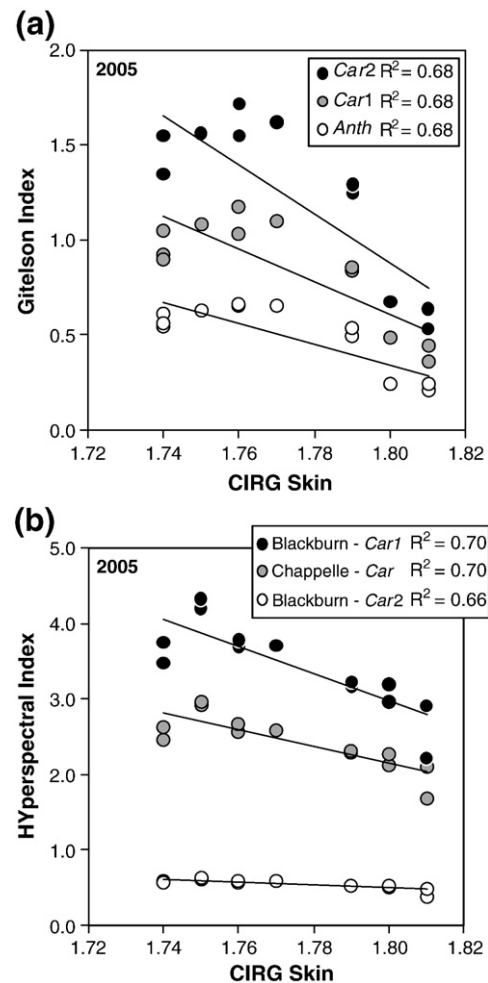


Fig. 13. Relationships between berry skin CIRG and physiological indices sensitive to Car and Anth pigment content.

the study) revealed the ability of physiological hyperspectral remote sensing indices to assess grape quality in vineyards affected by iron deficiency chlorosis, showing superior results compared to traditional vegetation indices. In particular, and despite their extensive use in remote sensing studies, NDVI and TCARI/OSAVI, utilized for vegetative vigor and leaf chlorophyll content retrieval, respectively, showed low coefficients of determination for physiological parameters linked to grape quality. In addition, results obtained from physiological indices sensitive to Car and Anth content underlined their potential for grape quality assessment, particularly phenolic composition and chromatic features of grapes (Table 8). Consistent results were obtained for the CIRG index, yielding coefficients of determination of $r^2 = 0.68$ (Gitelson Car2 index) and $r^2 = 0.56$ (Gitelson-Anth index) for the 2004 dataset, and $r^2 = 0.93$ (Gitelson-Car2 index) and $r^2 = 0.94$ (Gitelson-Anth index) for the 2005 dataset. Further results showed the Gitelson-Car2 index capability for IMAD quality index estimation with coefficients of determination of $r^2 = 0.72$ and 0.42 for the 2004 and 2005 datasets, respectively. In addition, berry skin quality parameters such as total polyphenol content (PT) and the CIRG index were well estimated through physiological indices, obtaining consistent relationships: $r^2 = 0.63$ (Chappelle-Car index) and $r^2 = 0.7$ (Blackburn-Car1 index) for PT and CIRG, respectively.

Phenolic compounds, anthocyanins and tannins are responsible for the color and astringency of red wines (Ribéreau-Gayon & Glories, 1987). Berry skin phenolic composition and color play a key role in quality determination for grapes such as Tempranillo. In

addition, the phenolic potential of grapes is of great importance in the Ribera del Duero *Appellation d'Origine* area, where other technologic quality parameters (sugar content, acidity, etc.) are not considered limiting factors for winemaking. In vineyards affected by iron chlorosis, the lack of photosynthetic pigments in leaves is a major factor limiting grape ripening. Previous work over the same area monitored in this study (Martín et al., 2007) reported that differences in grape quality parameters can be predicted by differences in leaf chlorophyll content of vines. The most obvious characteristic of the leaves from iron deficient plants is chlorosis, due to low concentration per area of chlorophylls and carotenoids. However, not all photosynthetic pigments are decreased to the same extent by iron deficiency, xanthophylls being less affected than chlorophylls and β -carotene (Abadia et al., 1999; Bertamini et al., 2001; Morales et al., 1994). On the other hand, an increase in leaf anthocyanin contents may reflect different biotic and abiotic stresses in plants, including mineral deficiencies (Chalker-Scott, 1999; Harbone, 1976).

The strong negative relationships reported between physiological indices based on Anth and Car with phenolic potential parameters, where relationships with indices based on chlorophyll content were not significant (Tables 6 and 7), suggest that an increase in the red component of color (derived from carotene and anthocyanin) in leaves in summer could be a better indicator than chlorosis to detect difficulties in phenolic ripening for vines affected by iron deficiency chlorosis.

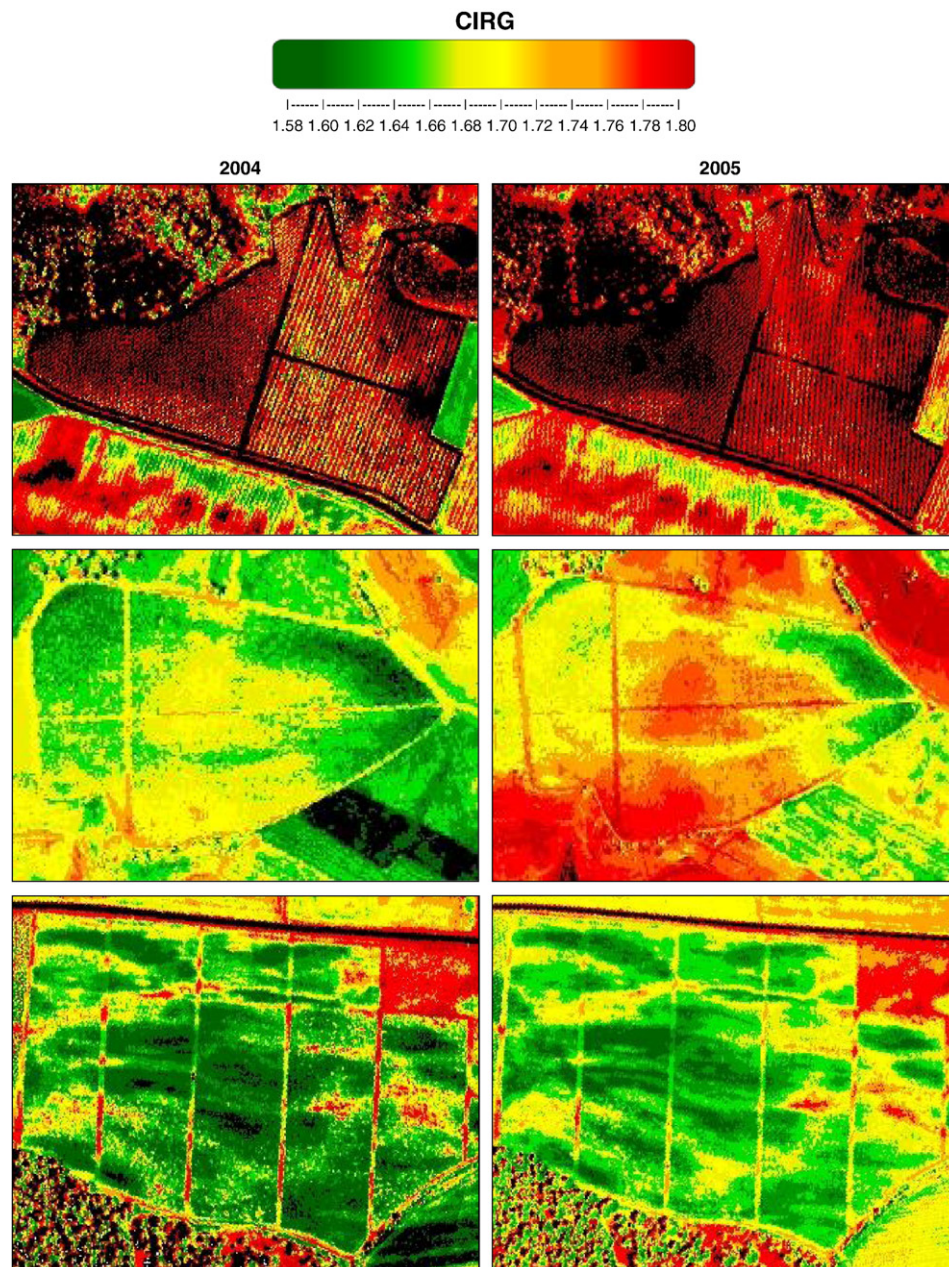


Fig. 14. Maps of Chromatic Index for Red Grapes (CIRG) obtained through the relationship with the Gitelson-Car2 physiological index for the 2004 and 2005 growing seasons.

Physicochemical conditions in calcareous soils, such as high pH, decrease the assimilation of micronutrients such as Cu and Zn, as well as iron (Martín et al., 2008). Our study showed significant relationships between K, Cu, Zn and B petiole levels with polyphenol contents in grapes and with *Car* and *Anth* physiological remote sensing indices. Therefore, these physiological spectral indices could be useful to detect nutritional effects related to iron deficiency affecting grape phenolic ripening.

5. Conclusions

This study suggests the potential of physiological remote sensing indices related to carotene and anthocyanin pigments content in leaves as grape quality indicators in vineyards affected by iron chlorosis, using high-resolution hyperspectral imagery. These indices could be used, as an alternative to others based on C_{ab} foliar content,

to monitor iron and other concurrent micronutrient deficiencies in vines before veraison, predicting the potential grape quality at harvest. The generation of quality maps from hyperspectral remote sensing would help assess within- and inter-field ripening variability by proposing spatially segmented harvesting zones for optimum winemaking.

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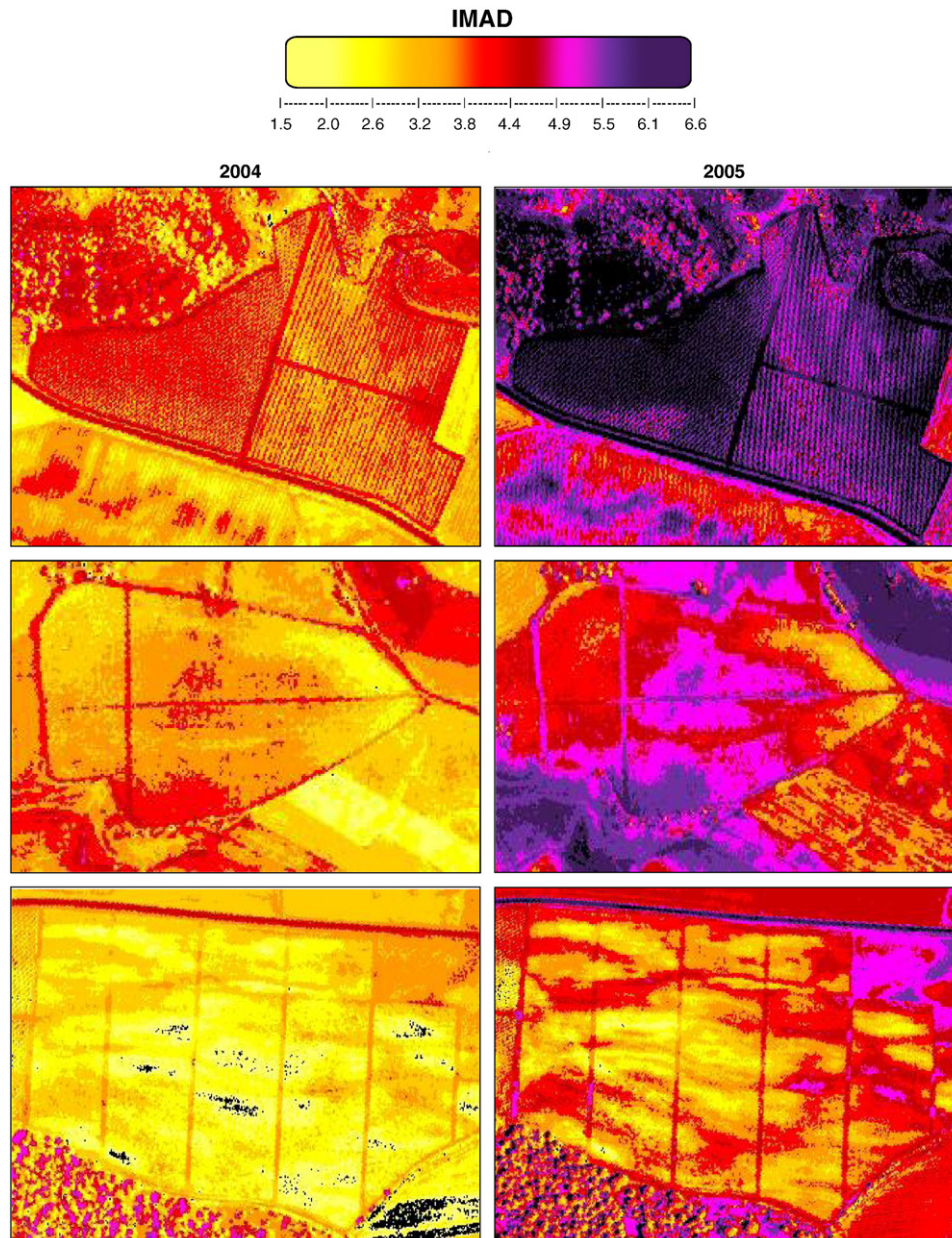


Fig. 15. Maps of IMAD ($^{\circ}\text{Vol}/\text{total acidity}$) obtained through the relationship with the Gitelson-Car2 physiological index for the 2004 and 2005 growing seasons.

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